

SABRAO Journal of Breeding and Genetics  
 58 (3) 1061-1072, 2026  
<http://doi.org/10.54910/sabrao2026.58.3.11>  
<http://sabraojournal.org/>  
 pISSN 1029-7073; eISSN 2224-8978



## PHYLOGENETIC ANALYSIS OF WILD GAMBIR (*UNCARIA* SPP.) AND DIVERSIFICATION OF ITS CULTIVATED SPECIES (*UNCARIA GAMBIR* [W. HUNTER] ROXB.)

SYAMSUARDI<sup>1\*</sup>, Y.S.W. MANUHARA<sup>2</sup>, N. WATHONI<sup>3</sup>, A. TAUFIQ<sup>1</sup>, and Z. AUDINA<sup>1</sup>

<sup>1</sup>Department of Biology, Faculty of Mathematics and Natural Sciences, Andalas University, Indonesia

<sup>2</sup>Department of Biology, Faculty of Science and Technology, Airlangga University, Indonesia

<sup>3</sup>Department of Pharmaceutics, Faculty of Pharmacy, Padjajaran University, Indonesia

\*Corresponding author's email: [syamsuardi@sci.unand.ac.id](mailto:syamsuardi@sci.unand.ac.id)

Email addresses of co-authors: [yosephine-s-w-m@fst.unair.ac.id](mailto:yosephine-s-w-m@fst.unair.ac.id), [nasrul@unpad.ac.id](mailto:nasrul@unpad.ac.id),

[ahmadtaufiq@sci.unand.ac.id](mailto:ahmadtaufiq@sci.unand.ac.id), [zikkraudina06@gmail.com](mailto:zikkraudina06@gmail.com)

### SUMMARY

The genus *Uncaria* (Rubiaceae) is a group of widespread woody plants with optimum diversity. Indonesia has over 103 cultivars of gambir and its wild relatives; however, research on wild *Uncaria*, particularly in West Sumatra, Indonesia, remains limited. The related study aimed to analyze the phylogenetic relationship of *Uncaria* spp. in West Sumatra, Indonesia, and recognize the speciation processes in *U. gambir* as an important germplasm. In this study, the collection of 16 gambir samples came from Pesisir Selatan (Siguntur and Taratak Tempatih) and Lima Puluh Kota (Simpang Kapuak and Ampalu), with 12 samples sequenced using data from the GenBank. Extracting DNA used the CTAB method, amplified with ITS markers, and sequenced. Carrying out the analysis engaged the Bioedit application for alignment and MEGA for phylogenetic reconstruction and genetic distance. The ITS sequence was 623 bp long with a GC content of 61.5%. *U. gambir* forms a monophyletic clade, found closely associated with the species *U. yunnanensis* and *U. lanosa*. The genetic distance (0.099–0.102) between *U. gambir* and other species was greater than between its own species (0.000–0.003). These results will support the domestication and selection of *U. gambir* for sustainable breeding and future conservation.

**Keywords:** *Uncaria* spp., wild relatives, cultivated species, domestication, ITS, phylogenetic analysis, genetic distance, germplasm conservation, West Sumatra

Communicating Editor: Prof. Naqib Ullah Khan

Manuscript received: November 25, 2025; Accepted: April 22, 2026.

© Society for the Advancement of Breeding Research in Asia and Oceania (SABRAO) 2026

**Citation:** Syamsuardi, Manuhara YSW, Wathoni N, Taufiq A, Audina Z (2026). Phylogenetic analysis of wild gambir (*Uncaria* spp.) and diversification of its cultivated species (*Uncaria gambir* [W. Hunter] Roxb.). *SABRAO J. Breed. Genet.* 58 (3) 1061-1072. <http://doi.org/10.54910/sabrao2026.58.3.11>.

**Key findings:** *Uncaria gambir* constitutes a remarkably homogeneous genetic group characterized by the least intraspecific variation. Its close relationship with the species *U. yunnanensis* and *U. lanosa* revealed an evolutionary connection and possible ancestral origin in East Asia. The results support sustainable breeding efforts and future conservation of *Uncaria* germplasm.

## INTRODUCTION

The flowering plant genus *Uncaria*, also known as Rubiaceae, is a polytypic genus comprising 34 species. Twenty-nine species existed in Asia and Australia, with three species observed in Africa and Madagascar and two species domesticated in the Americas (Ridsdale, 1978; Turner, 2018). Around 103 gambir cultivars and their wild relatives succeeded in their recognition in Indonesia. Each genotype has specific morphological characteristics, such as leaf shape, fruit shape, seed shape, taste, and fruit aroma (Ruwaida *et al.*, 2009). With numerous *Uncaria* species, Indonesia has emerged as the center of biodiversity along with its considerable distribution worldwide.

Sumatra is the main center of distribution, with seven species of *Uncaria* spp. found in the Sumatra Island (Nurainas *et al.*, 2020). Ridley (1922) reported five species of *Uncaria* spp. existing in West Sumatra, spread across almost all the districts, such as Padang, Agam, Tanah Datar, Sawah Lunto Sijunjung, Lima Puluh Kota, Pesisir Selatan, and Solok, Indonesia. It is a well-known fact that *Uncaria* spp. contain various complex and diverse secondary metabolites (Liang *et al.*, 2020; Wardi *et al.*, 2024), with the most common being the alkaloids and tannins (Munggari *et al.*, 2022). The said species has extensive uses in traditional medicine. However, to date, the community is only aware of the cultivated gambir (*Uncaria gambir*), while knowledge of wild gambir is rare and unexplored for its potential utilization and future conservation.

On *Uncaria* species, little research has taken place in West Sumatra. However, the current research on gambir has mostly focused on the use of cultivated gambir (*Uncaria gambir*) in West Sumatra (Wardi *et al.*, 2024), with less research being conducted on its wild relatives. The wild gambir species have the potential for secondary metabolites; however,

their exploration has not fully progressed. Exploration of wild gambir (*Uncaria* spp.) and its relatives is lacking; therefore, it necessitates sustainable conservation and utilization in traditional medicine.

At present, human activities, such as illegal logging, forest conversion, agricultural and plantation activities, and urbanization, have increased pressure on the tropical forests of Sumatra, Indonesia (Tsujino *et al.*, 2016; Eddy *et al.*, 2021), which seemed to be the habitat of wild gambir (Pinto *et al.*, 2022). In the long term, these conditions could threaten various plant species, including wild relatives of gambir, which could lead to the extinction of various wild *Uncaria* species, even before knowing their potential. This underscores the need for significant efforts to conserve germplasm for their sustainable utilization. Phylogenetic analysis of *Uncaria* species has grave consequences for sustainable use of these valuable plant genetic resources and future conservation in West Sumatra.

By understanding the evolutionary relationships among the *Uncaria* species, it becomes possible to identify centers of diversity and prioritize their conservation efforts in areas with high phylogenetic endemism. Additionally, phylogenetic data can help choose the right germplasm for growing and breeding purposes, which will help keep *Uncaria* resources available in the future. Molecular techniques using DNA barcoding approaches, such as ITS, *MatK*, and *trnL-F*, appeared to be crucial for phylogenetic studies of crop plants in West Sumatra, especially the wild gambir and its relatives (Vijayan and Tsou, 2010; Li *et al.*, 2015; Tnah *et al.*, 2019). Several molecular markers have been applicable to examine the phylogenetic relationship of the selected taxa in West Sumatra (Syamsuardi *et al.*, 2018; Huda *et al.*, 2019; Nuratika *et al.*, 2020; Audina *et al.*, 2025).

Therefore, the study based on exploration of the overall diversity of gambir will be helpful in effective conservation and sustainable utilization of these plant genetic resources. Moreover, phylogenetic analysis can help resolve taxonomic uncertainties and enable accurate identification, thereby facilitating the implementation of targeted bioprospecting strategies for plant diversity, particularly wild gambir. To this end, the following study aimed to determine the phylogenetic relationship of *Uncaria* spp. and analyze the diversification process of cultivated gambir indigenous to West Sumatra, Indonesia.

## MATERIALS AND METHODS

### Plant material

The collected *Uncaria* spp. totaled 16 from several locations in West Sumatra, Indonesia, such as Pesisir Selatan Regency (Taratak Tempatih and Siguntur) and Lima Puluh Kota Regency (Ampalu). The samples consisted of *U. homomalla*, *U. longiflora*, *U. lanosa*, and *U. cordata* and three samples each of the *U. gambir* landraces Udang, Cubadak, Riau Besar, and Riau Kecil. Furthermore, 12 sequences came from the GenBank, namely, *U. macrophylla*, *U. yunnanensis*, *U. lancifolia*, *U. laevigata*, *U. rhynchophylla*, *U. scandens*, *U. homomalla*, *U. hirsuta*, *U. sessilifructus*, *U. sinensis*, *U. lanosa*, and *Morinda citrifolia* (outgroup) (Table 1). The detailed view of sample collection locations appears in Figure 1. The germplasm collection comprised wild and cultivated gambir from West Sumatra. The detailed information on the gambir germplasm used in this study is available in Table 2.

### DNA extraction, amplification, and sequencing

DNA extraction, as carried out, used the CTAB method following the protocol of Doyle and Doyle (1987). The performance of DNA amplification utilized ITS primers (ITS4, 5'TCCTCCGCTTATTGATATGC3' and ITS5,

5'GGAAGTAAAAGTCGTAACAAGG3'), as described by White *et al.* (1990). The preparation of PCR reactions consisted of Biorun MyTaq Red Mix, which underwent the following thermal conditions: an initial pre-denaturation at 95 °C for one minute, followed by 15 s of denaturation at 95 °C, 15 s of annealing at 48.8 °C, and 10 s of extension at 72 °C. The amplification's completion had a final extension step at 72 °C for 10 min. The PCR products' analysis employed electrophoresis before visualization using a Gel Doc system. The conducted sequencing of PCR products used a single-pass DNA sequencing method at Apical Scientific Laboratory, Malaysia.

### Data analysis

Data analysis succeeded in processing the forward and reverse sequences obtained from the sequencing results. Initially, both sequences entailed merging into contigs to produce consensus sequences. Performing the alignment process then utilized BioEdit to ensure the accuracy of nucleotide comparisons for subsequent analyses (Ohied and Al-Badran, 2020). The use of MEGA later calculated genetic distances and constructed phylogenetic trees (Audina *et al.*, 2025). Based on molecular data, phylogenetic analysis is an important approach in understanding the relationship among species. This study utilized ITS markers obtained from nuclear DNA for analysis. The said method provided a more profound understanding of the evolutionary relationship and genetic diversity among the studied *Uncaria* spp. populations.

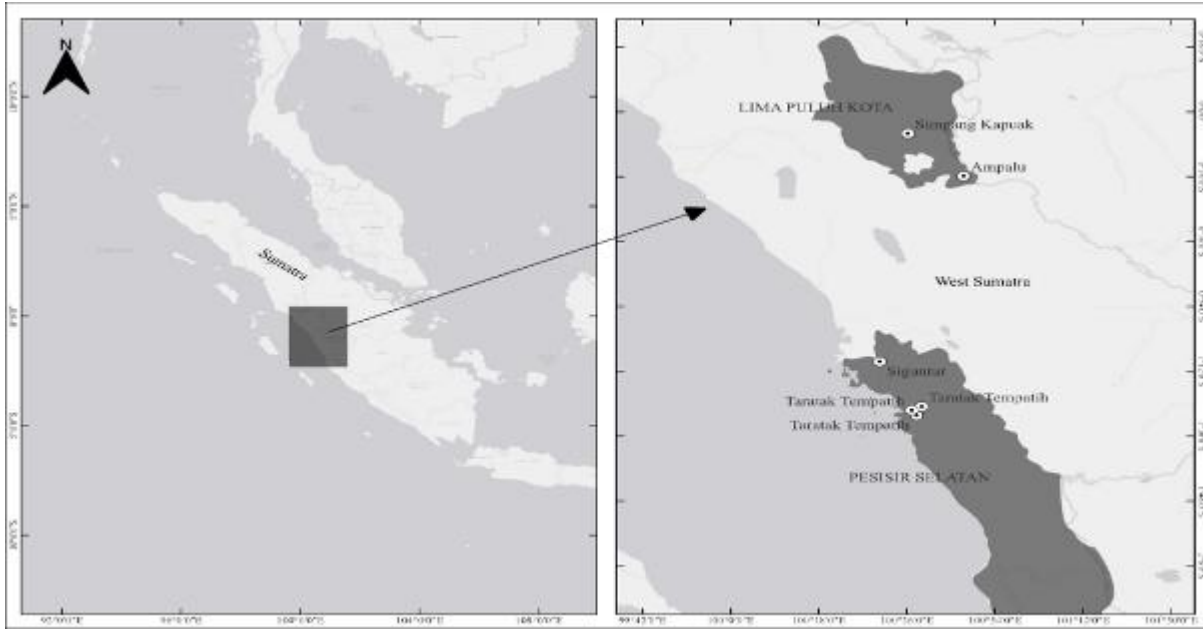
## RESULTS AND DISCUSSION

### Gambir speciation and phylogenetic analysis

After alignment, the length of the *Uncaria* spp. sequence was 623 bp. Based on the alignment results, the G-C content ranged from 61% to 62.8%, with an average of 61.5% (Tables 1 and 2). The G-C makes the DNA structure

**Table 1.** Genebank data used in data analysis.

Sample	Accession	Definition	G-C Content (%)	Description
<i>Uncaria macrophylla</i>	MF0333303	<i>U. macrophylla</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 26S ribosomal RNA gene, partial sequence.	62.8	Wild Gambir
<i>Uncaria yunnanensis</i>	KF881281.1	<i>U. yunnanensis</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	62.5	Wild Gambir
<i>Uncaria lancifolia</i>	KF881263.1	<i>U. lancifolia</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	62.2	Wild Gambir
<i>Uncaria laevigata</i>	KF881270.1	<i>U. laevigata</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	62.4	Wild Gambir
<i>Uncaria rhynchophylla</i>	KP092803.1	<i>U. rhynchophylla</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	61.8	Wild Gambir
<i>Uncaria scandens</i>	KF881275.1	<i>U. scandens</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	61.8	Wild Gambir
<i>Uncaria homomalla</i>	KF881251.1	<i>U. homomalla</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	61.5	Wild Gambir
<i>Uncaria hirsuta</i>	KM057050.1	<i>U. hirsuta</i> internal transcribed spacer 1, partial sequence; 5.8S ribosomal RNA gene, complete sequence; and internal transcribed spacer 2, partial sequence.	61.7	Wild Gambir
<i>Uncaria sessilifructus</i>	MT263757.1	<i>U. sessilifructus</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 26S ribosomal RNA gene, partial sequence.	62.2	Wild Gambir
<i>Uncaria sinensis</i>	FJ980386.1	<i>U. sinensis</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence	61.0	Wild Gambir
<i>Uncaria lanosa</i>	KC737635.1	<i>U. lanosa</i> internal transcribed spacer 1, partial sequence; 5.8S ribosomal RNA gene, complete sequence; and internal transcribed spacer 2, partial sequence.	62.5	Wild Gambir
<i>Morinda citrifolia</i>	FJ907061.1	<i>Morinda citrifolia</i> 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence.	62.9	Outgroup



**Figure 1.** Map of gambir (*Uncaria* spp.) sampling locations in West Sumatra, Indonesia.

**Table 2.** Distribution of gambir samples used in this study.

Sample	Accession Numbers	G-C Content (%)	Location	Description
<i>Uncaria gambir</i> (Landrace of Riau Kecil_1)	PZ274730	62.5	Siguntur (Pesisir Selatan)	Cultivated Gambir
<i>Uncaria gambir</i> (Landrace of Riau Kecil_2)	PZ274731	62.5		
<i>Uncaria gambir</i> (Landrace of Riau Kecil_4)	PZ274732	62.5		
<i>Uncaria gambir</i> (Landrace of Udang_1)	PZ274733	62.7		
<i>Uncaria gambir</i> (Landrace of Udang_3)	PZ274734	62.7		
<i>Uncaria gambir</i> (Landrace of Udang_4)	PZ274735	62.7		
<i>Uncaria gambir</i> (Landrace of Cubadak_1)	PZ274724	62.5		
<i>Uncaria gambir</i> (Landrace of Cubadak_2)	PZ274725	62.6		
<i>Uncaria gambir</i> (Landrace of Riau Besar_2)	PZ274727	62.7		
<i>Uncaria gambir</i> (Landrace of Riau Besar_3)	PZ274728	62.6		
<i>Uncaria gambir</i> (Landrace of Riau Besar_4)	PZ274729	62.6		
<i>Uncaria gambir</i> (Landrace of Cubadak_5)	PZ274726	62.5	Simpang Kapuak (Lima Puluh Kota)	Cultivated Gambir
<i>Uncaria homomalla</i>	PZ274736	62.2	Taratak Tempatih (Pesisir Selatan)	Wild Gambir
<i>Uncaria longiflora</i>	PZ274722	62.4		
<i>Uncaria lanosa</i>	PZ274723	62.6		
<i>Uncaria cordata</i>	PZ274737	62.0	Ampalu (Lima Puluh Kota)	Wild Gambir

more thermodynamically stable because it has three hydrogen atoms, while A-T (adenine-thymine) only has two (Smarda *et al.*, 2014).

In past studies, the frequent employment of ITS markers prevailed in plant taxonomy and phylogenetic research because

of their relatively rapid evolution and high resolution at the species level (Baldwin *et al.*, 1995; Alvarez and Wendel, 2003). By examining the ITS region, the determination of phylogenetic relationships among the *Uncaria* spp. species can succeed through the

similarities and differences in nucleotide sequences among the analyzed germplasm. By using the neighbor-joining (NJ) method, the phylogenetic analysis constructs a phylogenetic tree based on genetic distances formulated from nucleotide differences (Saitou and Nei, 1987).

Based on the phylogenetic analysis, the obtained cladogram showed the kinship among the germplasm of *Uncaria gambir* and several wild relatives. The cladogram revealed an informative evolutionary pattern regarding the origin and divergence pattern of the different species. The *Uncaria gambir* samples originating from numerous locations in West Sumatra, Indonesia (landrace of Udang, Cubadak, Riau Kecil, and Riau Besar), form a monophyletic clade and become separated from other *Uncaria* species. Past studies also revealed the monophyletic structure enunciated that *U. gambir* is a lineage sharing a common ancestor and has undergone differentiation and separation from its closest relatives (Ridsdale, 1978; Wardi *et al.*, 2023).

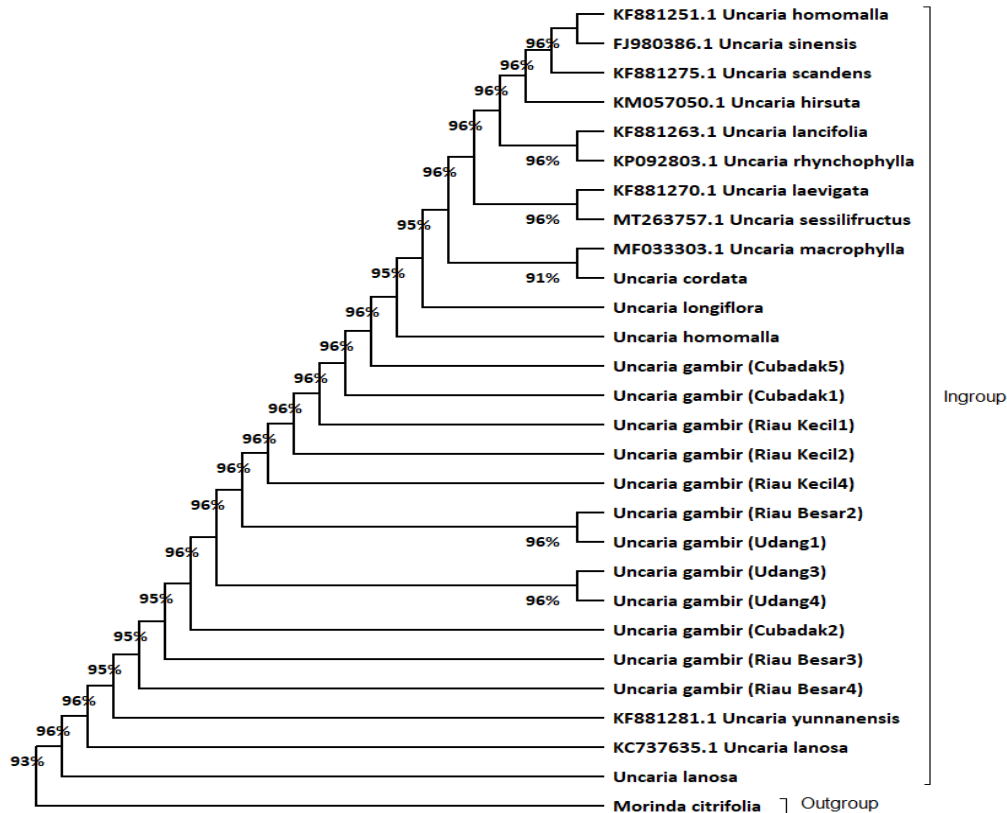
The proximity of the *U. gambir* clade to *U. yunnanensis* illustrated an obvious evolutionary relationship. The gambir species *U. yunnanensis* location was typically in China, whereas *U. gambir* is prevalent in the Sundaland Region, especially in Western Sumatra, Indonesia. This phylogenetic relationship indicated that the ancestor of *U. gambir* likely originated in mainland Southeast Asia, China. The divergence observed in the branches within the *U. gambir* clade revealed intraspecific genetic differentiation. The species *U. gambir* is one of the wild plants that have undergone domestication due to extensive cultivation by the community. The population selection with the highest catechin content and the main bioactive compounds in gambir plant extracts has led to artificial selection for certain morphological and biochemical traits (Munggari *et al.*, 2022). As a result, some locally cultivated cultivars of *U. gambir* showed phenotypic differences for leaf size as compared with wild populations. Although the said process has not formed a new species in the biological sense, this type of domestication reinforces practical reproductive isolation because the cultivated populations were

maintained vegetatively and the gene flow to wild populations was rare (Dempewolf *et al.*, 2012; Denham *et al.*, 2020).

Phylogenetic analysis based on the ITS marker yields high bootstrap values (91%–96%). If the bootstrap value is between 70 and 100, the possibility of branch changes decreases, indicating that the resulting clade structure has a high level of consistency. Conversely, if the bootstrap value is less than 70, the likelihood of changes in the clade structure increases (Rosidiani *et al.*, 2013). The phylogenetic tree successfully grouped all the *Uncaria* samples into one main group, separating them from the species *Morinda citrifolia* as an outgroup. The species *M. citrifolia* was an option for outgroup depending on its taxonomic proximity, as both belong to the family Rubiaceae but belong to different genera. The position of the species *M. citrifolia* at the bottom of the tree revealed this species was genetically distant from the genus *Uncaria*, proving it effective for rooting the phylogenetic tree (Figure 2) (Baldwin *et al.*, 1995).

The phylogenetic tree showed that several diverse *Uncaria* species formed a few separate groups. In particular, the species *U. gambir* group (obtained from Riau Kecil, Riau Besar, Udang, and Cubadak) created a single group, revealing these genotypes as genetically very close to each other and sharing a common ancestor. The cladogram places the species *U. gambir* separately from other gambir species in the genus *Uncaria*. These are *U. macrophylla*, *U. cordata*, *U. laevigata*, *U. senssilifructus*, *U. lancifolia*, *U. rhynchophylla*, *U. hirsuta*, *U. scandens*, *U. homomalla*, *U. sinensis*, *U. longiflora*, and *U. lanosa*. *U. gambir* proved closely associated with *U. yunnanensis* and *U. lanosa*, indicating that the ancestor of *U. gambir* originated in mainland Southeast Asia, China.

The species *U. macrophylla*, *U. cordata*, *U. laevigata*, *U. senssilifructus*, *U. lancifolia*, *U. rhynchophylla*, *U. hirsuta*, *U. scandens*, *U. homomalla*, and *U. sinensis* formed a single group. It indicates these species have relatively smaller genetic distances, ranging from 0.010 to 0.094, because of their close genetic relationship. Several other species' pairings in one clade,

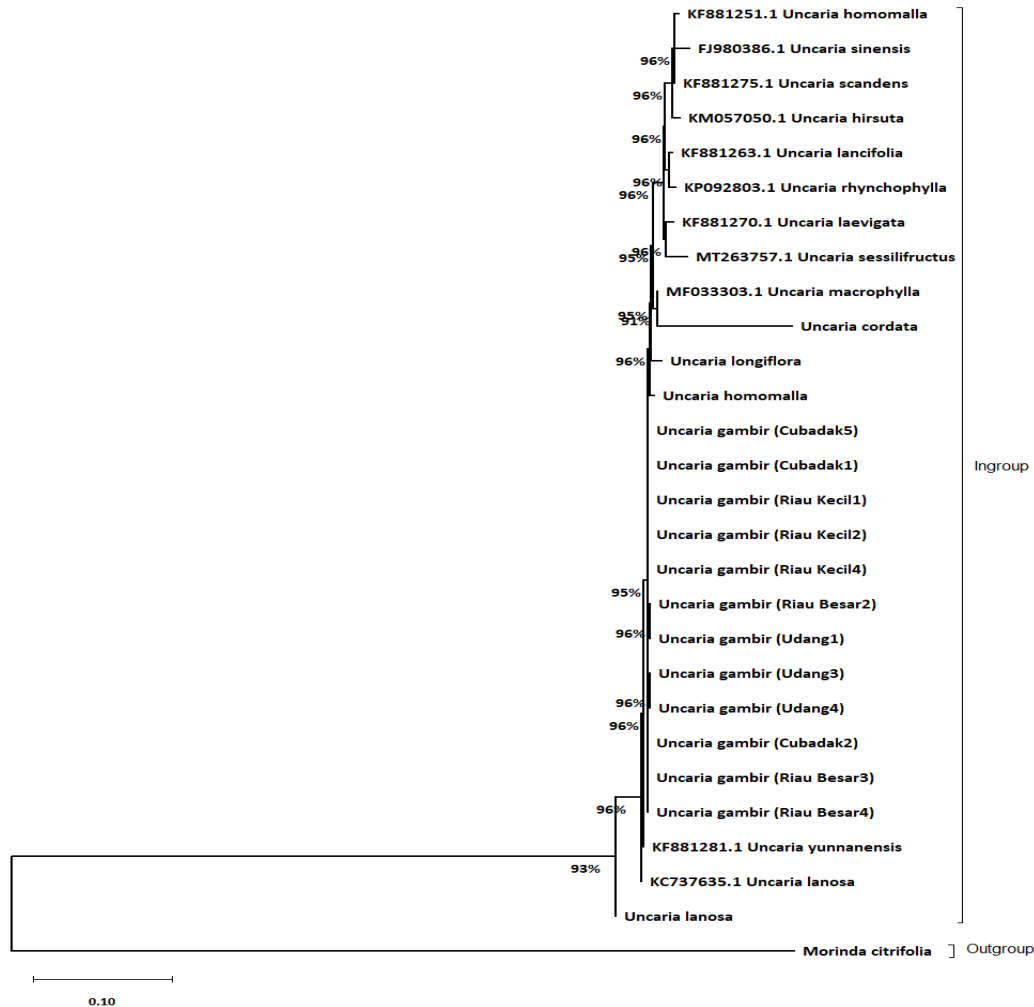


**Figure 2.** Phylogenetic tree of *Uncaria* spp. based on ITS markers using the neighbor-joining method.

i.e., *U. cordata* and *U. macrophylla*, *U. laevigata* and *U. sessilifructus*, and *U. lancifolia*, and *U. rhynchophylla*, showed similarities in their nucleotide base composition. Based on the tree created using the neighbor-joining method, one concludes that species *U. cordata* has the longest branching compared with other species (Figure 3). Thus, it became known that this species has a longer evolutionary journey than the other species. Conversely, a short branch reflects a relatively shorter evolutionary journey, as observed in the species *U. gambir*, which has the shortest branch length. In the phylogenetic tree, the branch length provides the evolutionary journey of the different species. The length of the branch indicates the duration of the species' evolutionary journey; a longer branch signifies a more extended evolutionary process, while a shorter branch denotes a more concise evolutionary trajectory (Tabatabaee *et al.*, 2023). This tendency

occurred to be associated with a smaller number of nucleotide base genetic variations (Liu *et al.*, 2022). Additionally, the *U. homomalla* sequence obtained from GenBank differs from the sequence generated from field samples. This difference is likely due to intraspecific genetic variation, whereby individuals of the same species but from different geographic locations may undergo genetic divergence as a result of population isolation, local selection, and genetic drift. This is consistent with the findings of Coissac *et al.* (2016), who stated that intraspecific variation in plants can lead to a decrease in sequence similarity, particularly in high-resolution markers.

In this classification, *U. gambir* belonged in group III along with the species *U. longiflora*, while the species *U. cordata* emerged in group I, and the species *U. lanosa* was in group IV. The phylogenetic analysis based on the cladogram showed that some of



**Figure 3.** Phylogenetic tree of *Uncaria* spp. using the neighbor-joining method.

these groups form consistent clusters; however, others revealed significant differences. In the phylogenetic tree, the species *U. cordata* appeared separate from the species *U. gambir* group, while close to the *U. macrophylla* species. This information was consistent with the taxonomic classification that places the species *U. cordata* and *U. macrophylla* in group I. In the taxonomic list, the traditional classification of the genus *Uncaria* showed seven major groups (Groups I-VII) (Turner, 2018).

In morphological classification, species *U. lanosa* was in group IV, while in the phylogenetic tree, it appeared in a separate cluster from the species *U. gambir*. This evidence supports that species *U. lanosa* was

genetically different. Genetic evidence thus supports the traditional morphological grouping. Nonetheless, certain sections require revisions in accordance with the genetic data (Liu *et al.*, 2022). This comparison implied that morphology-based classification alone may not fully reflect the true evolutionary relationship between the *Uncaria* species. For example, the species *U. gambir* and *U. longiflora* were separated in phylogenetic grouping, whereas in morphological grouping, these two species belonged to group III (Turner, 2018). Therefore, a study highly recommended the integration of morphological, genetic, and phylogenetic data to obtain an accurate and valid evolutionary classification (Villalobos-Cid *et al.*, 2020).

**Table 3.** Genetic distance of *Uncaria* spp.

Speci	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	
1																													
2	0.01																												
3	0.01	0.02																											
4	0.01	0.02	0.01																										
5	0.02	0.02	0.00	0.01																									
6	0.01	0.02	0.01	0.01	0.01																								
7	0.02	0.02	0.01	0.01	0.01	0.00																							
8	0.02	0.02	0.01	0.01	0.02	0.00	0.01																						
9	0.02	0.03	0.02	0.02	0.02	0.02	0.03	0.03																					
10	0.03	0.03	0.01	0.02	0.02	0.01	0.01	0.01	0.03																				
11	0.01	0.00	0.02	0.02	0.02	0.02	0.02	0.02	0.03	0.03																			
12	0.01	0.01	0.02	0.02	0.02	0.02	0.03	0.03	0.03	0.03	0.01																		
13	0.00	0.00	0.01	0.02	0.01	0.02	0.02	0.02	0.02	0.03	0.03	0.00	0.00																
14	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.02	0.03	0.00	0.01	0.00															
15	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00														
16	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00													
17	0.00	0.00	0.01	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00												
18	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00										
19	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00									
20	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00								
21	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00							
22	0.00	0.00	0.01	0.01	0.02	0.01	0.02	0.02	0.02	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00						
23	0.00	0.00	0.01	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00					
24	0.00	0.00	0.01	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00				
25	0.00	0.00	0.01	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00			
26	0.00	0.00	0.01	0.02	0.02	0.01	0.02	0.02	0.03	0.02	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
27	0.09	0.10	0.10	0.11	0.11	0.11	0.11	0.11	0.12	0.12	0.10	0.10	0.09	0.09	0.09	0.09	0.09	0.09	0.09	0.09	0.09	0.09	0.09	0.10	0.10	0.10	0.10		
28	1.02	1.00	1.02	1.02	1.01	1.03	1.05	1.01	1.03	1.05	1.00	1.01	0.97	1.01	1.00	1.01	1.01	1.01	1.00	1.00	1.01	1.01	1.01	1.01	1.01	1.01	1.01	1.03	1.33

Description: 1= MF033303.1 *U. macrophylla*, 2= KF881281.1 *U. yunnanensis*, 3= KF881263.1 *U. lancifolia*, 4= KF881270.1 *U. laevigata*, 5= KP092803.1 *U. rhynchophylla*, 6= KF881275.1 *U. scandens*, 7= KF881251.1 *U. homomalla*, 8= KM057050.1 *U. hirsuta*, 9= MT263757.1 *U. sessilifructus*, 10= FJ980386.1 *U. sinensis*, 11= KC737635.1 *U. lanosa*, 12= *U. longiflora*, 13= *U. lanosa*, 14= *U. gambir* (Cubadak1), 15= *U. gambir* (Cubadak2), 16= *U. gambir* (Cubadak5), 17= *U. gambir* (Riau Besar2), 18= *U. gambir* (Riau Besar3), 19= *U. gambir* (Riau Besar4), 20= *U. gambir* (Riau Kecil1), 21= *U. gambir* (Riau Kecil2), 22= *U. gambir* (Riau Kecil4), 23= *U. gambir* (Udang1), 24= *U. gambir* (Udang3), 25= *U. gambir* (Udang4), 26= *U. homomalla*, 27= *U. cordata*, and 28= *Morinda citrifolia* (Outgroup).

**Genetic distance analysis**

Genetic distance analysis served to evaluate the degree of kinship and genetic differences among the studied individuals. Based on the genetic distance, the values ranged from 0.000 to 0.003, which indicates the highest genetic relationship between *U. gambir* populations (Table 3). The smaller the genetic distance, the closer

the kinship of the species; conversely, the greater the genetic distance, the more distant the species kinship (Resida *et al.*, 2023). These results were greatly analogous to the neighbor-joining phylogenetic tree, indicating all the *U. gambir* species form a monophyletic clade, confirming all populations originate from the same ancestor without significant genetic divergence (Nei and Kumar, 2000).

Conversely, the genetic distance between *U. gambir* and other species in the genus *Uncaria* was greater than that between *U. gambir* and other *U. gambir* species, such as *U. sinensis* (0.030), *U. hirsuta* (0.023), and *U. rhynchophylla* (0.021). These values revealed a fairly high level of interspecies genetic divergence, consistent with their morphological and geographical distribution differences. The species *U. sinensis* and *U. hirsuta* existed in East Asia (China and Vietnam), while *U. gambir* was evident in the Western Southeast Asia (Sumatra and the Malay Peninsula). These considerable differences reinforce the hypothesis that geographic isolation played a vital role in this evolution (Coyne and Orr, 2004). Thus, the combination of NJ cladogram results and ITS genetic distances confirmed that *U. gambir* was distinctly a separate taxon from other *Uncaria* species but also remarkable with low intraspecific variation.

## CONCLUSIONS

The results showed species *U. gambir* forms a separate monophyletic clade with low genetic distance (0.000–0.003), confirming robust genetic uniformity and obvious separation from other *Uncaria* species. The greater genetic distance between *U. gambir* and other species, such as *U. sinensis*, *U. hirsuta*, and *U. rhynchophylla* (0.007–0.032), revealed significant interspecific divergence. Furthermore, the close phylogenetic relationship between the species *U. gambir*, *U. yunnanensis*, and *U. lanosa* (0.003–0.005) suggested a common evolutionary origin. Overall, this study found that the generated ITS-based molecular evidence provides a basis for refining the taxonomy of *Uncaria*.

## ACKNOWLEDGMENTS

The authors would like to thank the institution, Research and Community Service (LPPM), Andalas University, for its financial support of this research, with contract number 10/UN16.19/PT.01.03/RKI/2025.

## REFERENCES

- Alvarez I, Wendel JF (2003). Ribosomal ITS sequences and plant phylogenetic inference. *Mol. Phylogenet. Evol.* 29(3): 417–434. [https://doi.org/10.1016/S1055-7903\(03\)00208-2](https://doi.org/10.1016/S1055-7903(03)00208-2).
- Audina Z, Syamsuardi S, Mildawati M, Nurainas N, Christy P (2025). Haplotype diversity of *Uncaria gambir* (W. Hunter) Roxb. landraces in West Sumatra, Indonesia, based on internal transcribed spacer (ITS) markers. *SABRAO J. Breed. Genet.* 57(5): 2024–2035. <https://doi.org/10.54910/sabrao2025.57.5.23>.
- Baldwin BG, Sanderson MJ, Porter JM, Wojciechowski MF, Campbell CS, Donoghue MJ (1995). The ITS region of nuclear ribosomal DNA: A valuable source of evidence on angiosperm phylogeny. *Ann. Missouri Bot. Gard.* 82(2): 247. <https://doi.org/10.2307/2399880>.
- Coissac E, Hollingsworth PM, Lavergne S, Taberlet P (2016). From barcodes to genomes: Extending the concept of DNA barcoding. *Mol. Ecol.* 25: 1423–1428. <https://doi.org/10.1111/mec.13549>.
- Coyne JA, Orr HA (2004). *Speciation*. Sinauer, Sunderland.
- Dempewolf H, Hodgins KA, Rummell SE, Ellstrand NC, Rieseberg LH (2012). Reproductive isolation during domestication. *Plant Cell* 24(7): 2710–2717. <https://doi.org/10.1105/tpc.112.100115>.
- Denham T, Barton H, Castillo C, Crowther A, Dotte-Sarout E, Florin SA, Pritchard J, Barron A, Zhang Y, Fuller DQ (2020). The domestication syndrome in vegetatively propagated field crops. *Ann. Bot.* 125(4): 581–597. <https://doi.org/10.1093/aob/mcz212>.
- Doyle J, Doyle J (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19: 11–15.
- Eddy S, Milantara N, Basyuni M (2021). Carbon emissions as impact of mangrove degradation: A case study on the Air Telang protected forest, South Sumatra, Indonesia (2000–2020). *Biodiversitas* 22(4): <https://doi.org/10.13057/biodiv/d220464>.
- Huda M, Syamsuardi, Nurainas, Murni P, Maulidah R (2019). Genetic divergence landrace of langsung (*Lansium parasiticum*) from Siberut island based on ITS and *MatK* markers. *Indian J. Agric. Res.* 53(3): 338–342 <https://doi.org/10.18805/A-398>.

- Li X, Yang Y, Henry RJ, Rossetto M, Wang Y, Chen S (2015). Plant DNA barcoding: From gene to genome. *Biol. Rev.* 90(1): 157–166. <https://doi.org/10.1111/brv.12104>.
- Liang JH, Wang C, Huo XK, Tian XG, Zhao WY, Wang X, Sun CP, Ma XC (2020). The genus *Uncaria*: A review on phytochemical metabolites and biological aspects. *Fitoterapia* 147: 104772. <https://doi.org/10.1016/j.fitote.2020.104772>.
- Liu GQ, Lian L, Wang W (2022). The molecular phylogeny of land plants: Progress and future prospects. *Diversity* 14(10): 782. <https://doi.org/10.3390/d14100782>
- Munggari IP, Kurnia D, Deawati Y, Julaeha E (2022). Current research of phytochemical, medicinal and non-medicinal uses of *Uncaria gambir* Roxb.: A review. *Molecules* 27(19): 6551. <https://doi.org/10.3390/molecules27196551>.
- Nei M, Kumar S (2000). *Molecular Evolution and Phylogenetics*. Oxford University Press, New York.
- Nurainas N, Amolia RR, Taufiq A, Handika H, Syamsuardi S (2020). Flora of Sumatra: Vascular plant collection from Batang Toru forest deposited in ANDA Herbarium. Universitas Andalas, Padang, Indonesia.
- Nuratika E, Aseny N, Syamsuardi S, Nurainas N, Fitmawati F, Friadi F (2020). Clarification of Sumatran mulberry (*Morus macroura* Var. *Macroura*, Moraceae) from West Sumatra, Indonesia using nucleus ribosomal ITS (internal transcribed spacer) gene. *Indian J. Agric. Res.* 54(5): 635–640. <https://doi.org/10.18805/IJARE.A-508>.
- Ohied BM, Al-Badran AI (2020). Mitochondrial DNA (hypervariable region I) diversity in Basrah population-Iraq. *Genomics* 112(5): 3560–3564. <https://doi.org/10.1016/j.ygeno.2020.04.004>.
- Pinto GHT, Lopes AA, De Freitas Morel LJ, Crevelin EJ, Miranda CES, Contini SHT, De Castro Franca S, Bertoni BW, Pereira AMS (2022). Genetic diversity among genotypes of *Uncaria guianensis* (Aubl.) J.F. Gmel. maintained in an in vitro germplasm bank. *3 Biotechnol.* 12(1): 8. <https://doi.org/10.1007/s13205-021-03016-y>.
- Resida E, Chikmawati T, Ariyanti N, Fitmawati (2023). *Mangifera kemanga* Blume (Anacardiaceae) taxonomic assessment for genetic diversity based on molecular substantiation. *SABRAO J. Breed. Genet.* 55(1): 175–186. <https://doi.org/10.54910/sabrao2023.55.1.17>.
- Ridley NH (1922). *The Flora of Malay Peninsula*. L. Reeve & Co, London.
- Ridsdale CE (1978). A revision of *Mitragyna* and *Uncaria* (Rubiaceae). *Blumea* 24(1): 43–100.
- Rosidiani EP, Arumingtyas EL, Azrianingsih R (2013). Analisis variasi genetik *Amorphopallus muelleri* Blume dari berbagai populasi di Jawa timur berdasarkan sekuen intron trnL. *Floribunda* 4(6): 129–137.
- Ruwaida IP, Supriyadi S, Parjanto P (2009). Variability analysis of sukun durian plant (*Durio zibethinus*) based on RAPD marker. *Nus Biosci.* 1(2): 84–91. <https://doi.org/10.13057/nusbiosci/n010206>.
- Saitou N, Nei M (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4(4): 406–425. <https://doi.org/10.1093/oxfordjournals.molbev.a040454>.
- Smarda P, Bures P, Horova L, Leitch IJ, Mucina L, Pacini E, Tichy L, Grulich V, Rotreklova O (2014). Ecological and evolutionary significance of genomic GC content diversity in monocots. *Proc. Natl. Acad. Sci.* 111(39): 4096–4102. <https://doi.org/10.1073/pnas.1321152111>.
- Syamsuardi, Chairul, Murni P (2018). Analysis of genetic impurity of an original cultivar duku (*Lansium parasiticum* (Osbeck.) K.C. Sahn & Bennet.) from Jambi, Indonesia using ITS and *matK* gene. *Int. J. Environ. Agric. Biotechnol.* 3(2): 441–446. <https://doi.org/10.22161/ijeab/3.2.16>.
- Tabatabaee Y, Zhang C, Warnow T, Mirarab S (2023). Phylogenomic branch length estimation using quartets. *Bioinformatics* 30(39): 185–193. <https://doi.org/10.1093/bioinformatics/btad22>.
- Tnah LH, Lee SL, Tan AL, Lee CT, Ng KKS, Ng CH, Nurul FZ (2019). DNA barcode database of common herbal plants in the tropics: A resource for herbal product authentication. *Food Control* 95: 318–326. <https://doi.org/10.1016/j.foodcont.2018.08.022>.
- Tsujino R, Yumoto T, Kitamura S, Djameluddin I, Darnaedi D (2016). History of forest loss and degradation in Indonesia. *Land Use Policy* 57: 335–347. <https://doi.org/10.1016/j.landusepol.2016.05.034>.
- Turner IM (2018). A revised conspectus of *Uncaria* (Rubiaceae). *Webbia* 73(1): 9–21. <https://doi.org/10.1080/00837792.2018.1445363>.
- Vijayan K, Tsou CH (2010). DNA barcoding in plants: Taxonomy in a new perspective. *Curr. Sci.* 99(11): 1530–1541.
- Villalobos-Cid M, Salinas F, Inostroza-Ponta M (2020). Total evidence or taxonomic

- congruence? A comparison of methods for combining biological evidence. *J. Bioinform. Comput. Biol.* 16(3). <https://doi.org/10.1142/S0219720020500407>.
- Wardi ES, Syukur S, Chaidir Z, Jamsari J (2024). Genotypic identification and catechin profiling of *Uncaria gambir* in West Sumatra, Indonesia. *Biodiversitas* 25(3): 1151–1158. <https://doi.org/10.13057/biodiv/d250330>.
- Wardi ES, Syukur S, Chaidir Z, Jamsari J, Nova B (2023). DNA barcoding for the discrimination of *Uncaria gambir* and its closely related species using internal transcribed spacer genes. *F1000 Res.* 11: 106. <https://doi.org/10.12688/f1000research.74254.1>.
- White TJ, Bruns T, Lee S, Taylor J (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: PCR Protocols. CA: Academic Press, San Diego, pp. 315–322. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>.