

SABRAO Journal of Breeding and Genetics
 56 (3) 963-972, 2024
<http://doi.org/10.54910/sabrao2024.56.3.6>
<http://sabraojournal.org/>
 pISSN 1029-7073; eISSN 2224-8978



BIOCHEMICAL COMPOSITION, NUTRITIONAL VALUES, AND CALORIE CONTENT OF *ALLIUM* SPECIES: A SYSTEMATIC REVIEW

**M.I. IVANOVA¹, A.A. BAIKOV^{1*}, E.M. GINS², V.K. GINS¹, A.I. KASHLEVA¹,
 M.S. GINS^{1,3}, S.M. MOTYLEVA^{4*}, V.F. PIVOVAROV¹, and N.V. SMUROVA⁵**

¹Federal State Budgetary Scientific Institution Federal Scientific Vegetable Center, VNISSOK,
 Moscow Region, Russia

²Russian Potato Research Center, Kraskovo, Moscow Region, Russia

³Peoples' Friendship University of Russia, Moscow, Russia

⁴Strogoorganic Online Gardening School, Moscow, Russia

⁵All-Russian Scientific Research Institute of Medicinal and Aromatic Plants, Moscow, Russia

*Corresponding authors' emails: physiol@inbox.ru, motyleva_svetlana@mail.ru

Email addresses of co-authors: ivanova_170@mail.ru, katya.888888@yandex.ru, v.gins@inbox.ru,
vniioh@yandex.ru, anirr@bk.ru, plant.physiol@yandex.ru, plant.physiol@ro.ru

SUMMARY

The latest review discusses the chief metabolites of the genus *Allium* L., which characterize their nutritional qualities. The publications presented in the leading databases, such as Web of Science, Scopus, PubMed, Science Direct, and NCBI bore analysis. The vital active phytochemicals of *Allium* species include ascorbic acid, fatty acids, and carbohydrates. Many believe these biologically active compounds can prevent cancer development, coronary heart disease, and atherosclerosis.

Keywords: *Allium* L., biochemical composition, nutritional values, calorie content, onions, chives, garlic, leeks

Key findings: The analysis of the experimental data on metabolites of onion crops based on their nutritional values in light of the functional nutrition concept is currently relevant. The genus *Allium* L. species are an integral part of the human diet and a valuable source of biologically active substances.

Communicating Editor: Dr. Kamile Ulukapi

Manuscript received: October 09, 2023; Accepted: March 06, 2024.

© Society for the Advancement of Breeding Research in Asia and Oceania (SABRAO) 2024

Citation: Ivanova MI, Baikov AA, Gins EM, Gins VK, Kashleva AI, Gins MS, Motyleva SM, Pivovarov VF, Smurova NV (2024). Biochemical composition, nutritional values, and calorie content of *Allium* species: a systematic review. *SABRAO J. Breed. Genet.* 56(3): 963-972. <http://doi.org/10.54910/sabrao2024.56.3.6>.

INTRODUCTION

The first introduction of the functional nutrition concept occurred in Japan around three decades ago. It refers to foods that contain components beneficial to one or more bodily functions relating to comfort and health and lessening the risk of different diseases (Roberfroid, 2000). Plant-derived compounds, defined as phytochemicals, produce various health-promoting effects when used. Presently, a primary aim of food researchers is to identify the evidence demonstrating the functionality of foods. Therefore, studies on the biological properties of putative phytochemicals and the biochemical composition of plant foods are highly appealing (Mamedov et al., 2017; Gins et al., 2020).

The genus *Allium* L., belonging to the family Amaryllidaceae, includes about 1200 species and has progressive replenishment (Herden et al., 2016). The most recent classification suggested 15 sub-genera and 56 sections for *Allium* (Brullo et al., 2019). The key centers of *Allium* diversity exist in Central Asia, the Mediterranean, and the Western part of North America (Friesen et al., 2006; Jabbes et al., 2011). The flora of Asia is abundant in onions. The regions abundant with more than 130 species appear in Western Asia (Turkey, Iran, and Afghanistan). However, Central Asia has more than 190 designated species. Non-Asian onion types account only for 38.2%. The natural growth and cultivation environment exists only in the South by the tropics and in the North by the 60th parallel. *Allium* species are mainly prevalent in temperate, semi-arid, and arid regions of the Northern Hemisphere (Fritsch and Friesen, 2002).

The genus *Allium* L. often attracts researchers' attention given its valuable medicinal, nutritional, and decorative properties, adaptive capabilities, resistance to pests and diseases, and ecological plasticity. These characteristics contribute to the competitiveness of species and the manifestation of a high degree of acclimatization outside natural areas and broad geographical allocation (Friesen et al., 2006). Humans consume more than 40 wild onion

species, some of which serve to develop new types of cultivars and hybrids (Teshika et al., 2019).

Members of the genus *Allium* L. contain most of human nutrition's essential components. Ascorbic acid, chlorophylls, and secondary metabolites (carotenoids and phenolic compounds) mainly characterized the onion leaves' food and pharmacological values. The nutritional values comprise beneficial properties of a product that can satisfy the daily physiological needs of the human body with specific substances and the release of energy due to oxidation of the consumed components. The nutritional value directly depends on the biochemical composition of the products with deliberations concerning its consumption in accepted quantities of the chief organic substances, i.e., proteins, carbohydrates, fats, vitamins, and acids.

Detailed data on the daily human needs are available in Table 1. In onion family crops, the calorie content varies from 1.3% (tubular green leaves of the onions) to 9.71% (garlic bulbs) of the daily human needs. In this case, the protein contents also ranged from 1.2% (onion bulbs) to 7.07% (greens of garlic), carbohydrates without dietary fibers ranged from 1.36% (green chives) to 22.14% (garlic bulbs), and food fibers ranged from 4.0% (garlic greens) to 12.5% (green chives) of the daily human requirements. Fat content rated at 0.15%–1.04%, and water (2.16%–3.41%) of the daily human demands.

The nutritional structure of the genus *Allium* species is very complex and difficult to appraise. Plant metabolite levels principally depended upon the genetic and environmental factors and transport and storage conditions. Growth factors, such as light, temperature, humidity, soil type, fertilization, microbial and insect damage, UV stress, heavy metals, and pesticides amend the composition of plant metabolites (Orcutt and Nilsen, 2000).

Over the past decade, a considerable sum of data has accumulated on the identification, biochemical characterization, localization, and health benefits of *Allium* metabolites. The presented review is an attempt to summarize information on the

Table 1. Calorie content and nutritional values of onion crops (per 100 g FW) and percentage in the daily human need.

<i>Allium</i> species	Caloric content	Proteins	Fats	Carbohydrates	Dietary fiber	Water
<i>A. fistulosum</i> (leaves)	2.2	2.07	0.6	2.93	12	3.34
<i>A. cepa</i> (leaves)	1.3	1.41	0.15	2.29	6	3.41
<i>A. cepa</i> (bulb)	2.61	1.2	0.15	5.43	8.5	3.26
<i>A. porrum</i> (leaves)	2.35	2.17	0.3	4.5	11	3.23
<i>A. schoenoprasum</i> (leaves)	1.96	3.59	1.04	1.36	12.5	3.34
<i>A. ascalonicum</i> (bulb)	4.69	2.72	0.15	9.71	16	2.93
<i>A. ursinum</i> (leaves)	2.28	2.61	0.15	4.36	5	3.26
<i>A. sativum</i> (leaves)	3	7.07	0.15	3.71	4	2.93
<i>A. sativum</i> (cloves)	9.71	6.96	0.75	22.14	10.5	2.16

USDA National Nutrient Database for Standard Reference: <https://data.nal.usda.gov/dataset/usda-national-nutrient-database-standard-reference-legacy-release>.

metabolites that determine the nutritional values of the genus *Allium* and their vital role in the human daily diet. The primary content levels and ranges of variation of the foremost metabolites in onion crops attained analysis.

Inclusion criteria

This review solely focused on analyzing the biochemical composition of onion crops, including biologically active compounds of pharmacological significance. The review included the Web of Science, Scopus, PubMed, Science Direct, SciFinder, and Google Scholar databases. The online sources used were the Research Gate, National Center for Biotechnology Information (NCBI), Springer Nature Open Access, and Wiley Online Library.

Biochemical composition

Onions contain numerous phytochemicals, with organosulfur compounds as the most crucial. These compounds add the odor and taste characteristics and contain most of their biological possessions. Another essential group of biologically active compounds is polyphenols, which include phenolic acids and flavonoids, responsible for the bulbs' color trait. The edible parts of the onion are rich in carbohydrates, mainly glucose, and fructose, while the outer scales of the bulbs contain significant amounts of galactose and arabinose. Essential amino acids (arginine and glutamic acid), as relevant nitrogen stores, also contribute to the nutritional values of onion

species. The onions also contain several other complex bioactive components, such as saponins, vitamins (A, C, B6, and B9), and minerals (P, K, Ca, Mg, Zn, Mn, Na, Fe, Br, I, Se, and Cu).

The organic compounds found in the species of genus *Allium* are a product of primary and secondary plant metabolism. Primary metabolites, i.e., carbohydrates, amino acids, fatty acids, and organic acids, contribute to the growth and development, respiration and photosynthesis, and hormone and protein synthesis. Primary metabolites are prevalent in all the species within extensive phylogenetic groups and proceed to form using the same biochemical pathways. Secondary metabolites, viz., flavonoids, carotenoids, sterols, phenolic acids, alkaloids, and glucosinolates, determine the color of vegetables, protect plants from herbivores and microorganisms, attract pollinators and seed dispersers, and act as indicator molecules under stress conditions (Crozier *et al.*, 2006).

Carbohydrates

In vegetables, the carbohydrates come in the form of monosaccharides (glucose, fructose, arabinose, galactose, and rhamnose), disaccharides (sucrose, maltose, and trehalose), sugar alcohols (sorbitol, mannitol, and xylitol), oligosaccharides (raffinose, fructooligosaccharides, and galactooligosaccharides), and polysaccharides (starch, cellulose, hemicellulose, and pectin). On their physiological and nutritional role, the

classification of carbohydrates is often available and unavailable. Available carbohydrates are those that enzymes hydrolyze in the human gastrointestinal system into monosaccharides, such as sucrose and digestible starch. Monosaccharides do not necessitate digestion and can continue direct absorption into the bloodstream and defend the cell's protein-lipid components, participating in the detoxification of free radicals (Siddiqui et al., 2020).

Unavailable carbohydrates (sugar alcohols, many oligosaccharides, and non-starch polysaccharides) are nonhydrolyzable by endogenous human enzymes, with microorganisms fermenting them in the large intestine before absorption (Asp, 1996). Fructooligosaccharides and non-starch polysaccharides are vital components of dietary fiber. Plant components described as dietary fiber typically include non-starch polysaccharides, resistant oligosaccharides, lignin, and related substances, such as resistant starch, waxes, cutin, and suberin (De-Vries, 2003). These materials pass through the gastrointestinal tract as fibers, where colonic microorganisms modify and digest them (Blaut, 2002). The high-fiber foods consumption has proven to reduce the symptoms of chronic constipation, diverticular disease, and several types of colitis (Stollman and Raskin, 2004). Therefore, a conclusion surfaced that a low-fiber diet may increase the risk of colon cancer, cardiovascular diseases, and obesity (Marlett, 2001; Slavin, 2005; Vernia et al., 2021).

Fructooligosaccharides (fructans) assemble in the species of *Allium* in different forms (Shiomi et al., 2005). Considerable information emerged about polysaccharides, including those based on fructose, in various species of the genus *Allium* (Shepherd and Gibson, 2006; Flores et al., 2016). This genus' most typical fructans are inulin, inulin neoseris, and levan neoseris (Livingston et al., 2009). In different species of *Allium*, the fructans content varies from 1.1 to 10.1 g/100 g, including in *A. sativum* L. (9 to 16 g/100 g), and the concentration of fructose and sucrose in different types of onions varies from 0.7% to 4.0% of fresh weight (Shepherd and Gibson,

2006). Levan-type fructans have a pronounced moisturizing effect on the skin, parallel to that of hyaluronic acid, as well as anti-inflammatory properties, and therefore, an endowing use in cosmetology (Kim et al., 2006).

In round *Allium* species (*A. aflatunense* B. Fedtsch., and *A. caeruleum* Pall.), the elevated molecular weight glucofructans (GFH) contents are significantly higher (22.22% to 41.40%) than in the group of bulbous-rhizomatous plants with large rhizomes and poorly developed bulbs (*A. ledebourianum* Schult. & Schult. F., *A. nutans* L., and *A. ramosum* L.). The group of bulbous-rhizomatous plants with large bulbs and poorly developed rhizomes (*A. obliquum* L., *A. altaicum* Pall., and *A. strictum* Schrad.) occupied an intermediate position between the two previous groups, which are similar significantly. In this group, the average high molecular weight glucofructans was 20.34% ± 3.64%. The varied accumulation of low molecular weight (GFL) and high molecular weight glucofructans in different plant organs is evident. In *A. altaicum* and *A. obliquum* (bulb-rhizomatous species with well-developed tubers), the bulb's GFL and GFH contents were 1.5–2.0 times higher than the rhizomes.

In *A. nutans* and *A. ramosum* with small bulbs, the differences in the content of GFL are low, and in *A. ramosum* rhizomes, the GFH content was 1.3 times higher than in bulbs. The glucofructans accumulation significantly varies during the growing season, showing an association with different intensities of plant growth and development. The maximum accumulation of GFH was prominent during the flowering period; however, their concentration noticeably decreased by the end of the growing season. In different species, the GFL content reaches its maximum in varied phenophases, as determined by the peculiarities of the seasonal development of the species (Shiomi et al., 2005).

Pectin is a part of the cell tissue's structural elements of higher plants and acts as a binding and strengthening component of the cell wall and regulates water metabolism (Minzanova et al., 2021). The onion pectin displayed a bimodal molar mass distribution,

suggesting two polymer fractions with different molar masses, and it is vital in the observed promising emulsifying and emulsion stabilizing potential (Neckebroeck *et al.*, 2021).

Amino acids

Amino acids, as transitional products, are the structural components of proteins, playing an influential role in the metabolism of plants and animals. Proteins perform crucial functions at the tissue level and act as enzymes, hormones, and antibodies. Dietary proteins are the chief source of amino acids. Most proteins have enzymes breaking them down into amino acids for absorption in the small intestine. Humans can synthesize several amino acids, including alanine, asparagine, aspartic acid, cysteine, cystine, glutamic acid, glutamine, glycine, proline, serine, and tyrosine. The nine amino acids called essential must come from the diet, including histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, and valine (Hounsome *et al.*, 2018). Vegetables contain all the essential amino acids; however, some may be in smaller quantities than the amount required for humans (Young and Pellett, 1994).

Arginine (Arg) is an amino acid that appears to be sufficient in onions, supporting

the hypothesis that onions use Arg as a nitrogen reservoir, contributing to the nutritional value of the *Allium* species (Fredotovič *et al.*, 2017, 2020). In addition to Arg, glutamic acid (Glu) and lysine (Lys) occur in large quantities. Garlic cloves also contain a high content of arginine (Lawson and Gardner, 2005). Moreover, the main free amino acids found in garlic are glutamine (Gln), asparagine (Asn), Glu, and Lys (Lee and Harnly, 2005). Interestingly, the proline content in onion seeds is a reliable indicator of cultivar tolerance to water stress (Golubkina *et al.*, 2023).

In onions, the average data on the content of amino acids (mg/100 g FW) comprised, i.e., Arg - 104, Histidine - 14, Isoleucine - 14, Leucine - 25, Lys - 39, Methionine - 2, Phenylalanine - 25, Tryptophan - 14, and Valine - 21 (Hounsome *et al.*, 2018). The analyzed 17 bound amino acids and 38 free amino acids from the leaves of *A. victorialis* L. revealed the total content of component amino acids and free amino acids at 2693.28 mg/100 g and 535.39 mg/100 g (Cho *et al.*, 2011).

In *A. cornutum* Vis. and *A. cepa* L., the Arg content was 16.492 and 12.213 mg/g DW, respectively, while the Glu content was 14.964 and 11.238 mg/g DW, respectively (Table 2)

Table 2. Amino acid composition of *Allium* (Fredotovič, 2020).

Amino Acids	<i>A. cornutum</i> Vis. (mg/g DW)	<i>A. cepa</i> L. (mg/g DW)
Asp (Aspartic acid)	4.941 ± 0.043	6.100 ± 0.083
Ser (Serine)	2.921 ± 0.004	2.846 ± 0.031
Glu (Glutamic acid)	14.964 ± 0.008	11.238 ± 0.096
Gly (Glycine)	2.042 ± 0.013	2.217 ± 0.015
His (Histidine)	1.758 ± 0.015	1.667 ± 0.052
Arg (Arginine)	16.492 ± 0.256	12.213 ± 0.094
Thr (Threonine)	3.324 ± 0.013	4.003 ± 0.076
Ala (Alanine)	1.675 ± 0.002	2.027 ± 0.023
Pro (Proline)	1.434 ± 0.024	1.702 ± 0.128
Cys (Cysteine)	0.144 ± 0.002	0.023 ± 0.005
Tyr (Tyrosine)	1.719 ± 0.010	2.277 ± 0.049
Val (Valine)	2.005 ± 0.003	2.375 ± 0.072
Met (Methionine)	0.013 ± 0.005	0.010 ± 0.005
Lys (Lysine)	2.322 ± 0.010	2.328 ± 0.018
Ileu (Isoleucine)	1.426 ± 0.006	1.818 ± 0.007
Leu (Leucine)	2.284 ± 0.014	2.949 ± 0.026
Phe (Phenylalanine)	2.198 ± 0.014	2.378 ± 0.006

(Fredotovič et al., 2020). The amino acids Glu and Arg may be responsible for the color of onion seeds (Dini et al., 2008). In addition to its importance in human metabolism, free amino acids also affect the vegetable taste. Glycine and alanine are sweet; valine and leucine are bitter, and aspartic acid and glutamate taste sour and umami (Bachmanov et al., 2016; Wu, 2021). Prominently, in onions, these amino acids are potential fingerprints of geographical origin (Ianni et al. 2018).

Ascorbic acid

Ascorbic acid, as an antioxidant, performs the biological functions of a reducing agent and coenzyme of some metabolic processes. This compound protects against several reactive oxygen species types, i.e., free radicals and peroxides (Dzhos et al., 2023). It also synthesizes neurotransmitters, steroid hormones, and collagen, helps heal wounds and burns, prevents blood clotting and bruising, and strengthens the walls of human capillaries (Hounsoum et al., 2018).

In onions, the growth conditions and nitrogen fertilization affect the ascorbic acid content (Kazimierczak et al., 2021). In the studied onion crops (33 species) in the Moscow Region, Russia, the ascorbic acid content in the leaves varied from 119.2 (*A. pskemense* B. Fedtsch.) to 133.5 (*A. suworowii* Regel) and, on average (126.0 ± 4.1 mg/100g fresh weight) (Ivanova et al., 2019).

Fatty acids

Fatty acids are the prime components of fats, consisting of a hydrophilic carboxylate group attached to a long hydrocarbon chain. An 80% of the total lipids of all *Allium* species consisted of four fatty acids, i.e., linoleic (46%–53%), palmitic (20%–23%), oleic (4%–13%), and α -linolenic acid (3%–7%). In *A. cepa*, 43 fatty acids are distinct, where 18 are more than 0.4%, and four are more than 2.5%. In *A. sativum*, 70 fatty acids are noteworthy, 14 at above 0.4%, and only four are above 2.5%. The *A. porrum* L. determined 50 fatty acids, having 12 being more than 0.4% and four

being more than 2.5%. Phospholipids comprise a few specific fatty acids, while neutral lipids contain a wide range of fatty acids (Tsiaganis et al., 2006). *A. sativum* lipids amounted to 0.42% of raw plant tissue; the chief fatty acids are linoleic, palmitic, linolenic, and oleic acid (Tsiaganis et al., 2006).

In *A. fistulosum* L., the most common fatty acids in descending order of their content were linoleic (52.87%) > oleic (17.57%) > palmitic (9.80%) > stearic (8.81%) > linolenic (2.88%) > palmitoleic (2.84%) > myristic (1.28%) > behenic (1.23%). With a concentration of less than 1.00% are lauric, arachidonic, behenic, and lignoceric acids. Linoleic and oleic acids were the most abundant of all fatty acids, as well as the total unsaturated fatty acids, with the two making up 70.44% of the content of all fatty acids. Palmitic and stearic acids were the two most abundant saturated fatty acids, accounting for 18.61% of all fatty acids. Total unsaturated fatty acids (77.35%) dominate the total saturated fatty acids (22.63%), while the polyunsaturated fatty acids (56.34%) were much higher than that of monounsaturated (21.04%) (Adeyeye, 2020). The chief fatty acids detected in *A. sativum* were linoleic acid, which ranged from 32.54% to 58.97%, palmitic acid (ranging from 13.62% to 18.46%), and oleic acid (ranging from 6.21% to 14.74%) (Petropoulos et al., 2018).

In plant oil, the high level of essential fatty acids is an advantage in food consumption, and a better total unsaturated/saturated ratio makes this oil nutritionally applicable for domestic purposes. Linoleic acid, the most abundant fatty acid in oil, is also valuable as it prevents cardiovascular diseases (Singh et al., 2022). It helps relieve rough and flaky skin, maintaining smooth and silky skin (Adeyeye, 2020). Oleic acid also deters cardiovascular ailments (Arsic, 2020). Oleic acid can decrease plasma triacylglycerol and cholesterol concentrations without affecting plasma high-density lipoprotein cholesterol levels; linoleic and linolenic acids help reduce plasma concentrations of total cholesterol, low-density lipoprotein cholesterol, minute-density lipoprotein cholesterol, apolipoprotein B, and

apolipoprotein A-1, lower the blood pressure, and protect against coronary heart diseases.

Linoleic acid can reduce the levels of inflammation-related factors and has a significant ameliorating effect on both acute and chronic inflammation; α -linolenic and γ -linolenic acids possess anti-inflammatory and anti-allergic properties to improve the immune system, also serving as a precursor of the prostaglandin and tissue hormones. Additionally, linoleic acid helps inhibit cancer growth in the human breast, colon, skin, stomach, and leukemia (Tian *et al.*, 2023). Likewise, linolenic acid is crucial for nutrition, and it is an essential polyunsaturated fatty acid used in the biosynthesis of arachidonic acid. The available arachidonic acid also serves as a foundation for the biosynthesis of prostaglandin and thromboxane in humans (Majima *et al.*, 2023). Myristic and stearic acids benefit as surfactants, emulsifiers, and thickeners in cosmetics (Adeyeye, 2020). The vegetable oil *A. fistulosum* has an acceptable polyunsaturated/saturated (P/S) ratio of 3.42 (Adeyeye, 2020). A high P/S ratio reduces serum cholesterol and atherosclerosis, preventing heart diseases (Xue *et al.*, 2023).

Mineral elements

Mineral elements are structural components vital in enzyme activity, osmotic pressure control for cell turgor and growth, and acid-base and water-salt metabolism (Tetyannikov *et al.*, 2022). The green onion leaves are an excellent source of mineral elements that enter the human body as ions in balanced concentrations. In onion leaves, the contents of the mineral elements follow the order of Zn > Fe > Si > Na > P > Cl > Mo > Mg > S > Ca > Cu > K (Nemtinov *et al.*, 2020, 2021).

The *A. flavescens* Bess. accumulates five microelements (Cr, Ni, Cu, Zn, and Se) and contains high Fe, Co, and Mn concentrations. These features motivate the significant promise of this species to procure these microelements. The content of Cr in 100 g of leaves of this species equals 84% of the daily human need for this microelement. *A. fistulosum* L., *A. odorum* L., and the broad-

leaved form of *A. nutans* specifically amass the Cu, Zn, and Se. *A. montanum* Schmidt comprised Zn accretion. *A. angulosum* L. and *A. schoenoprasum* L. cause the accrual of Se. The broad-leaved forms of *A. schoenoprasum* L., *A. nutans* L., and *A. odorum* L. accumulate higher amounts of Zn and Cu than their narrow-leaved counterparts (Golubev *et al.*, 2003).

Garlic is a valuable source of macro- and microelements, especially potassium, calcium, magnesium, phosphorus, iron, manganese, selenium, vanadium, copper, and zinc. The garlic dry matter has about 4000 mg/kg of potassium (48.1% from the mass of all elements contained in cloves), 1500 mg/kg of phosphorus (18.3%), 1400 mg/kg of sulfur (17.2%), almost 900 mg/kg of calcium (10.7%), 300 mg/kg of magnesium (3.6%), 100 mg/kg of sodium (1.3%), 20 mg/kg (0.24%) of silicon, 16 mg/kg (0.20%) of iron, and 14 mg/kg (0.17%) of zinc. The content of manganese, aluminum, copper, and boron has the range of 1.8 mg/kg (0.02%)–6.9 mg/kg (0.084%); chromium, nickel, selenium, cadmium, cobalt, iodine, lithium, vanadium, germanium, and lead from 0.22 mg/kg (0.003%) to 0.013 mg/kg (0.0002%), and mercury - 0.0043 mg/kg (0.00005%).

Garlic cultivation in diverse climatic conditions leads to a significant change in the content of various elements. In this case, vanadium, boron, and cobalt contents can vary by 20 times, and the content of iron and germanium emerged more than 1000 times. In garlic, the variability of the elemental composition lessened based on the genotypes compared with the growing conditions. In general, the elemental composition depended upon cultivars (varies 1.1–4.7 times) while depending on growing conditions (2.1–57.4 times). Differences in iron content were according to cultivars (1.5 times) and on growing conditions (57.4 times), respectively, copper (1.4 and 54.0 times) and sodium (1.1 and 26.8 times). Garlic is a unique elemental complex, and according to indicators of sulfur, zinc, manganese, and copper, it corresponds to meat and fish (Polyakov *et al.*, 2020).

Phytoremediation

Phytoremediation has contaminants removed from the soil using crop plants. Soil phytoremediation from organic and inorganic pollutants is evident based on plant-microbial interactions, and the presence of microorganisms resistant to pollutants promoted plant growth in the plant rhizosphere (Muratova et al., 2023). Garlic has the potential to absorb and hyper-accumulate heavy metals. It was apparent that the accumulation of Cd and Pb was much higher with inter-planting (Chatterjee and Chatterjee, 2021; Hussain et al., 2021). The enrichment of heavy metals in onion farmland may induce the accumulation of heavy metals in onion and cause food safety issues (Kharisu et al., 2019).

CONCLUSIONS

Given their distinctive aroma and rich nutritional content, onions and garlic are highly desirable edible plants widely used in cooking. Their regular utilization can prevent hyperlipidemia, diabetes, hypertension, cardiovascular diseases, and inflammatory conditions. Onions and their by-products provide many health benefits but require more research to identify the underlying molecular mechanisms and their interaction. *Allium* species (especially garlic) may also pose a safety risk due to elevated heavy metals accumulation.

Funding

This work proceeded within the topic of the state task of the All-Russian Scientific Research Institute of Medicinal and Aromatic Plants № FGUU-2022-0014.

REFERENCES

Adeyeye A (2020). Physico-chemical characteristics and fatty acid profile of *Allium fistulosum* vegetable plant. *Int. J. Adv. Chem.* 8(2): 239-243.

- Arsic A (2020). Oleic acid and implications for the Mediterranean diet. In: V.R. Preedy and R.R. Watson (Eds.), *The Mediterranean Diet*, 2nd Edition. Academic Press, pp. 267-274.
- Asp NG (1996). Dietary carbohydrates: Classification by chemistry and physiology. *Food Chem.* 57(1): 9-14.
- Bachmanov AA, Bosak NP, Glendinning JI, Inoue M, Li X, Manita S, McCaughey SA, Murata Y, Reed DR, Tordoff MG, Beauchamp GK (2016). Genetics of amino acid taste and appetite. *Adv. in Nutr.* 7(4): 806S-822S.
- Blaut M (2002). Relationship of prebiotics and food to intestinal microflora. *Eur. J. Nutr.* 41: 111-116.
- Brullo S, Brullo C, Cambria S, del Galdo GG, Salmeri C (2019). *Allium albanicum* (Amaryllidaceae), a new species from Balkans and its relationships with *A. meteoricum* Heldr. & Hausskn. ex Halácsy. *PhytoKeys* 119: 117-136.
- Chatterjee S, Chatterjee S (2021). Phytoremediation of arsenic using *Allium sativum* L. as a model system. In: P.K. Shit, P.P. Adhikary, D. Sengupta (Eds.), *Spatial Modeling and Assessment of Environmental Contaminants. Environmental Challenges and Solutions*. Springer, Cham. pp. 121-136.
- Cho J-Y, Park Y-J, Oh D-M, Rhyu D-Y, Kim Y-S, Chon S-U, Kang S-S, Heo B-G (2011). Amino acid contents and various physiological activities of *Allium victorialis*. *Korean J. Plant Resour.* 24(2): 150-159.
- Crozier A, Clifford M, Ashihara H (2006). *Plant Secondary Metabolites: Occurrence Structure and Role in the Human Diet*. Oxford, UK: Blackwell Publishing Ltd.
- De-Vries JW (2003). On defining dietary fibre. *Proc. Nutr. Soc.* 62(1): 37-43.
- Dini I, Tenore GC, Dini A (2008). Chemical composition, nutritional value, and antioxidant properties of *Allium cepa* L. var. *tropeana* (red onion) seeds. *Food Chem.* 107(2): 613-621.
- Dzhos EA, Pyshnaya ON, Mamedov MI, Baikov AA, Gins MS, Tukuser YP, Matykina AA, Shafigullin DR, Gins EM, Motyleva SM (2023). Biologically active compounds transform during the ripening stages in greenhouse tomatoes. *SABRAO J. Breed. Genet.* 55(2): 533-540.
- Flores AC, Morlett JA, Rodríguez R (2016). Inulin potential for enzymatic obtaining of prebiotic oligosaccharides. *Crit. Rev. Food Sci. Nutr.* 56(11): 1893-1902.
- Fredotovič Ž, Soldo B, Šprung M, Marijanovič Z, Jerkovič I, Puizina J (2020). Comparison of organosulfur and amino acid composition

- between triploid onion *Allium cornutum* Clementi ex Visiani, 1842, and common onion *Allium cepa* L., and evidences for antiproliferative activity of their extracts. *Plants* 9(1): 98.
- Fredotovič Ž, Šprung M, Soldo B, Ljubenkovič I, Budič-Leto I, Bilušič T, Čikeš-Čulič V, Puizina J (2017). Chemical composition and biological activity of *Allium cepa* L. and *Allium cornutum* (Clementi ex Visiani 1842) methanolic extracts. *Molecules* 22(3): 448.
- Friesen N, Fritsch RM, Blattner FR (2006). Phylogeny and new intrageneric classification of *Allium* (Alliaceae) based on nuclear ribosomal DNA ITS sequences. *Aliso* 22(1): 372-395.
- Fritsch RM, Friesen N (2002). Evolution, domestication and taxonomy. In: H.D. Rabinowitch and L. Currah (Eds.), *Allium Crop Science: Recent Advances*. CABI, Wallingford, UK. pp. 5-30.
- Gins V, Fotev Y, Baikov A, Mizrukhnina Y, Gadzhikurbanov A, Rebouh Y (2020). Survey of antioxidants and photosynthetic pigments in the newly introduced crops of Russia: *Benincasa hispida*, *Vigna unguiculata*, *Cucumis metuliferus* and *Momordica charantia*. *Res. on Crops*. 21(2): 339-343.
- Golubev FV, Golubkina NA, Gorbunov YN (2003). Mineral composition of wild onions and their nutritional value. *Appl. Biochem. Microbiol.* 39: 532-535.
- Golubkina N, Romanova O, Romanov V, Krivenkov L, Shevchenko T, Murariu OC, Vecchiatti L, Hamburda SB, Caruso G (2023). Varietal differences of yield, morphological, and biochemical parameters of *Allium cepa* L. under precipitation excess in different phenological phases. *Stresses* 3(3): 541-554.
- Herden T, Hanelt P, Friesen N (2016). Phylogeny of *Allium* L. subgenus *Anguinum* (G. Don. ex W.D.J. Koch) N. Friesen (Amaryllidaceae). *Mol. Phylogenet. Evol.* 95: 79-93.
- Hounsborne N, Hounsborne B, Lobo MG (2018). Biochemistry of vegetables. In: M. Siddiq and M. Uebersax (Eds.), *Handbook of Vegetables and Vegetable Processing*. John Wiley and Sons, Hoboken, NJ. pp. 25-46.
- Hussain J, Wei X, Xue-Gang L, Shah SRU, Aslam M, Ahmed I, Abdullah S, Babar A, Jakhar AM, Azam T (2021). Garlic (*Allium sativum*) based interplanting alters the heavy metals absorption and bacterial diversity in neighboring plants. *Sci. Rep.* 11(1): 5833.
- Ianni F, Lisanti A, Marinozzi M, Camaioni E, Pucciarini L, Massoli A, Sardella R, Concezzi L, Natalini B (2018). Amino acid content in onions as potential fingerprints of geographical origin: The case of *Rossa da inverno* sel. Rojo Duro. *Molecules* 23(6): 1259-1271.
- Ivanova MI, Bukharov AF, Baleev DN, Bukharova AR, Kashleva AI, Seredin TM, Razin OA (2019). Biochemical composition of leaves of *Allium* L. species in the conditions of the Moscow region. *Achiev. Sci. Technol. AICis* 33(5): 47-50.
- Jabbes N, Geoffriau E, Le Clerc V, Dridi B, Hannechi C (2011). Inter simple sequence repeat fingerprints for assess genetic diversity of Tunisian garlic populations. *J. Agric. Sci.* 3(4): 77-85.
- Kazimierczak R, Średnicka-Tober D, Barański M, Hallmann E, Góralska-Walczak R, Koczyńska K, Rembiałkowska E, Górski J, Leifert C, Rempelos L, Kaniszewski S (2021). The effect of different fertilization regimes on yield, selected nutrients, and bioactive compounds profiles of onion. *Agronomy* 11(5): 883.
- Kharisu CS, Koki IB, Ikram R, Low KH (2019). Elemental variations and safety assessment of commercial onions (*Allium cepa*) by inductively coupled plasma-mass spectrometry and chemometrics. *Int. Food Res. J.* 26: 1717-1724.
- Kim KH, Chung CB, Kim YH, Kim KS, Han CS, Kim CH (2006). Cosmeceutical properties of levan produced by *Zymomonas mobilis*. *Int. J. Cosmetic Sci.* 28(3): 231.
- Lawson LD, Gardner CD (2005). Composition, stability, and bioavailability of garlic products used in a clinical trial. *J. Agric. Food Chem.* 53(16): 6254-6261.
- Lee J, Harnly JM (2005). Free amino acid and cysteine sulfoxide composition of 11 garlic (*Allium sativum* L.) cultivars by gas chromatography with flame ionization and mass selective detection. *J. Agric. Food Chem.* 53(23): 9100-9104.
- Livingston III DP, Hinch DK, Heyer AG (2009). Fructan and its relationship to abiotic stress tolerance in plants. *Cell. Mol. Life Sci.* 66(13): 2007-2023.
- Majima M, Hosono K, Ito Y, Amano H, Nagashima Y, Matsuda Y, Watanabe S, Nishimura H (2023). A biologically active lipid, thromboxane, as a regulator of angiogenesis and lymphangiogenesis. *Biomed. Pharm.* 163: 114831.
- Mamedov MI, Pishnaya ON, Baikov AA, Pivovarov VF, Dzhos EA, Matykina AA, Gins MS (2017). Antioxidant contents of pepper *Capsicum* spp. for use in biofortification. *Agric. Biol. (Sel'skokhozyaistvennaya biologiya)*. 52(5): 1021-1029.

- Marlett JA (2001). Dietary fiber and cardiovascular disease. In: S.S. Cho and M.L. Dreher (Eds.), *Handbook of Dietary Fiber*. Marcel Dekker Inc., New York, pp. 17-30.
- Minzanova ST, Mironov VF, Mindubaev AZ, Tsepaveva OV, Mironova LG, Milyukov VA, Gins VK, Gins MS, Kononkov PF, Babayev VM, Pivovarov VF (2021). Extraction and physicochemical characterization of pectin polysaccharides from amaranth leaves. *Agric. Biol. (Sel'skokhozyaistvennaya Biologiya)*. 56(3): 591-601.
- Muratova A, Gorelova S, Golubev S, Kamaldinova D, Gins M (2023). Rhizosphere microbiomes of *Amaranthus* spp. grown in soils with anthropogenic polyelemental anomalies. *Agronomy* 13(3): 759.
- Neckebroek B, Verkempinck SHE, Van Audenhove J, Bernaerts T, de Wilde d'Estmael H, Hendrickx ME, Van Loey AM (2021). Structural and emulsion stabilizing properties of pectin rich extracts obtained from different botanical sources. *Food Res. Int.* 141: 110087.
- Nemtinov V, Kostanchuk Y, Motyleva S, Katskaya A, Timasheva L, Pekhova O, Pashtetskiy V, Kulikov I, Medvedev S, Bokhan A (2020). Mineral composition of *Allium cepa* L. leaves of southern subspecies. *Potravinarstvo Slovak J. Food Sci.* 14: 216-223.
- Nemtinov VI, Kostanchuk YN, Pashtetskiy VS, Motyleva SM, Bokhan AI, Caruso G, Katskaya AG, Timasheva LA, Pekhova OA (2021). Biochemical and cytological features of onion bulbs and leaves collected from various ecogeographical origins. *SABRAO J. Breed. Genet.* 53(4): 543-560.
- Orcutt DM, Nilsen ET (2000). *The Physiology of Plants under Stress Soil and Biotic Factors*. New York: John Wiley and Sons Inc.
- Petropoulos SA, Fernandes Â, Ntatsi G, Petrotos K, Barros L, Ferreira ICF (2018). Nutritional value, chemical characterization and bulb morphology of Greek garlic landraces. *Molecules* 23: 319.
- Polyakov A, Alekseeva T, Muravieva I (2020). The elemental composition of garlic (*Allium sativum* L.) and its variability. *E3S Web Conf.* 175: 01016.
- Roberfroid MB (2000). Concepts and strategy of functional food science: The European perspective. *Am. J. Clin. Nutr.* 71: 1660S-1664S.
- Shepherd SJ, Gibson PR (2006). Fructose malabsorption and symptoms of irritable bowel syndrome: Guidelines for effective dietary management. *J. Am. Diet. Assoc.* 106(10): 1631-1639.
- Shiomi N, Benkeblia N, Onodera S (2005). The metabolism of the fructooligosaccharides in onion bulbs: A comprehensive review. *J. Appl. Glycosci.* 52: 121-127.
- Siddiqui H, Sami F, Hayat S (2020). Glucose: Sweet or bitter effects in plants - a review on current and future perspective. *Carbohydr Res.* 487: 107884.
- Singh RB, Watanabe S, Li D, Nakamura T, Juneja LR, Takahashi T, Wichansawakun S, Wilczynska A, Jantan I, Sulaeman A, Ridwan H, Bharadwaj K, Mojto V, Kartikey K, Rawal S, Smail MMA (2022). Effects of polyunsaturated fatty acid-rich diets and risk of non-communicable diseases. In: R.B. Singh, S. Watanabe, and A.A. Isaza (Eds.) *Functional Foods and Nutraceuticals in Metabolic and Non-Communicable Diseases*. Academic Press, pp. 165-185.
- Slavin JL (2005). Dietary fiber and body weight. *Nutrition* 21(3): 411-418.
- Stollman N, Raskin JB (2004). Diverticular disease of the colon. *Lancet* 363(9409): 631-639.
- Teshika JD, Zakariyyah AM, Zaynab T, Zengin G, Rengasamy KR, Pandian SK, Fawzi MM (2019). Traditional and modern uses of onion bulb (*Allium cepa* L.): A systematic review. *Crit. Rev. Food Sci. Nutr.* 59: S39-S70.
- Tetyannikov NV, Motyleva SM, Gins MS, Kozak NV, Panischeva DV, Mertvisheva ME, Kabashnikova LF, Domanskaya IN, Pilipovich TS (2022). Drought effects on mineral composition of the leaves and seeds of *Amaranthus tricolor* and *Amaranthus cruentus*. *SABRAO J. Breed. Genet.* 54(2): 426-436.
- Tian M, Bai Y, Tian H, Zhao X (2023). The chemical composition and health - promoting benefits of vegetable oils - A review. *Molecules* 28: 6393.
- Tsiaganis MC, Laskari K, Melissari E (2006). Fatty acid composition of *Allium* species lipids. *J. Food Compos. Anal.* 19: 620-627.
- Vernia F, Longo S, Stefanelli G, Viscido A, Latella G (2021). Dietary factors modulating colorectal carcinogenesis. *Nutrients* 13(1): 143.
- Wu G (2021). *Amino Acids: Biochemistry and Nutrition*. CRC Press, Boca Raton, FL.
- Xue L, Yang R, Wang X, Ma F, Yu L, Zhang L, Li P (2023). Comparative advantages of chemical compositions of specific edible vegetable oils. *Oil Crop Sci.* 8(1): 1-6.
- Young VR, Pellett PL (1994). Plant proteins in relation to human protein and amino acid nutrition. *Am. J. Clin. Nutr.* 59: 1203S-1212S.