

SABRAO Journal of Breeding and Genetics 55 (5) 1510-1525, 2023 http://doi.org/10.54910/sabrao2023.55.5.6 http://sabraojournal.org/ pISSN 1029-7073; eISSN 2224-8978



AEGILOPS L. GENETIC DIVERSITY IN SOUTHWESTERN REGION OF UZBEKISTAN

I. DJABBAROV¹, F. SOBIROV^{1,2*}, T. BOZOROV², K. TURAKULOV², and S. BABOEV²

¹Samarkand State University named after Sharof Rashidov, Samarkand City, Uzbekistan ²Institute of Genetics and Experimental Plant Biology, Academy of Science of the Republic of Uzbekistan, Tashkent, Uzbekistan *Corresponding author's email: faridun.sobirov@internet.ru

Email addresses of co-authors: djabborov59@mail.ru; tohirbozorov@yahoo.com, sai-baboev@yandex.ru

SUMMARY

As wheat donors, wild species of the genus Aegilops L. play a vital role in practical breeding to improve wheat production because of their strong relationship and wide genetic diversity. Using nine simple sequence repeat (SSR) markers helped assess the genetic diversity in 96 collected samples of four species belonging to the genus Aegilops, i.e., Aegilops tauschii Coss (D), Ae. cylindrica Host (CD), Ae. crassa Boiss (DDM), and Ae. triuncialis L. (UC). The said collection came from 21 sites of various expeditions located in three regions of Southwestern Uzbekistan (Samarkand, Urgut, Kitab, and Shakhrisabz regions). Generally, 102 distinct alleles were found, with an average of 11.33 alleles per primer. The total number of species-specific amplicons was 35. The polymorphism detected varied from 28.6% (for the WSP107 primer) to 77.0% (for the WSP130 and WSP192 primers). The mean values of polymorphism information content (PIC) and expected heterozygosity (Ho) for all samples were 0.675 and 0.527, respectively. Based on nine SSR markers, on average, the genetic distance indices (GD) varied from 0.63 to 0.77. The highest genetic similarity (GD = 0.77) recorded occurred between the species Ae. crassa and Ae. cylindrica, whereas the least (GD = 0.48), between Ae. cylindrica and Ae. triuncialis with their taxonomic classification. Genus Aegilops samples from the same region often attain an identical subgrouping, which might be due to relatedness by genetic parameters. The gene pool of native species of the genus Aegilops from the Southwestern region of Uzbekistan may provide suitable alleles for wheat improvement and adaptation in the future.

Keywords: *Aegilops* L., *Ae. tauschii, Ae. crassa, Ae. cylindrica, Ae. triuncialis,* genetic diversity, D-genome, SSR primers

Key findings: Results revealed from nine SSR markers a marker WSP513 was the most polymorphic. In the genus *Aegilops* L., the species *Ae. tauschii* showed the highest genetic diversity, and samples collected from Khazraty Bashi (21) enunciated the foremost polymorphism among the studied regions.

Communicating Editor: Prof. Ijaz Rasool Noorka

Manuscript received: February 1, 2023; Accepted: August 31, 2023. © Society for the Advancement of Breeding Research in Asia and Oceania (SABRAO) 2023

Citation: Djabbarov I, Sobirov F, Bozorov T, Turakulov K, Baboev S (2023). *Aegilops* L. genetic diversity in Southwestern Region of Uzbekistan. *SABRAO J. Breed. Genet.* 55(5): 1510-1525. http://doi.org/10.54910/sabrao2023.55.5.6.

INTRODUCTION

Aegilops is a large genus of the tribe Triticeae, including 23 annual species (11 diploid, eight tetraploid, and four hexaploid species) with diverse ploidy (Konstantinos and Bebeli, 2010). These Aegilops species have main distribution in the Mediterranean and Southwestern and Central Asia (Kilian et al., 2011), with centers of origin in Georgia, Armenia, Azerbaijan, and Northern Iran (Hammer et al., 2001; Van-Slageren, 1994), and Cyprus, Lebanon, Israel, Syria, Iraq, Southeast Turkey, Southwest Iran, and Northwest Jordan (Lelley et al., 2000). These diverse regions have high concentrations of the Aegilops species, which are also welladapted compared with other genera. Some Aegilops species even had involvement in the wheat evolution (Ae. tauschii - the D-genome donor and of great importance in the origin of common wheat); thus, potentially beneficial in wheat improvement programs (Konstantinos and Bebeli, 2010; Tilman et al., 2011; Reynolds et al., 2012; Ogbonnaya et al., 2013; Kishii, 2019; Abbas et al., 2020).

Aegilops and Gossipum species often have a wide variety of the desired traits characterizing them, which provides an invaluable gene pool for resistance to biotic and abiotic stressors in wheat breeding (Saghai-Maroof et al., 1984; Alnaddaf et al., 2012; Martynov et al., 2015; Arzani and Ashraf, 2017; Elbashir et al., 2017; Itam et al., 2020; Shavkiev et al., 2022, 2023; Makamov et al., 2023; Kurbanbaev et al., 2023). For cereal resistance to biotic stresses, 20% of genes were found in various species of the genus Aegilops (Roder et al., 1995). Several Aegilops species are also distinct with exceptional yield and quality traits, such as, growth, large ears, and the high content of microelements and gluten in the grains (Kavrakova, 2009; Buronov et al., 2023).

Molecular analysis can better suit the study of the genetic diversity of *Aegilops* species and facilitate the complex trait association and the selection of suitable donors for breeding purposes (Aliyev *et al.*, 2007; Hajiyev *et al.*, 2015). In this regard, DNA markers have breeders' wide use of these to assess the *Aegilops* intraspecific genetic

diversity. Like the SSR markers, molecular markers have proven the most effective because of their polymorphism, reproducibility, co-dominance, and simplicity (Roder et al., 1995). These markers are chromosomespecific, with an extensive use of breeders for identifying the helpful genes (Fufa et al., 2005), assessing genetic diversity (Ehtemam et al., 2010; Masoumi et al., 2012; Hajiyev et al., 2015; Henkrar et al., 2016; Vieira et al., 2016), and phylogenetic relationships in various crops, including wheat and its wild relatives (Liu et al., 2016; Luo et al., 2017; Abbasov et al., 2018). However, despite longtime research on the genetic diversity of Aegilops species, each species requires additional information on their distribution and diversity in different agroecological zones.

The southwest of Uzbekistan is a mountainous region within which various crop plant species are prevalent, including diverse species of the genus Aegilops (Sobirov and Djabbarov, 2021; Chorshanbiev et al., 2023). The area has a unique natural habitat for wild relatives of wheat, wherein five species of the genus *Aegilops* occupy a prominent place. This region has limited study of the Aegilops species on their genetic diversity and breeding utility. Given these facts, the local populations of Aegilops species should have much consideration as potential sources of helpful alleles for wheat improvement and adaptation programs.

Therefore, it seemed relevant to assess the genetic diversity of *Aegilops'* local species grown in different natural and climatic conditions of the southwestern region of Uzbekistan using SSR markers since it is the first time in this region that research on the genus *Aegilops* using SSR markers will happen. The presented study sought to assess the genetic diversity of the genus *Aegilops* collection comprising 96 specimens of four local species distributed in the southwestern region of Uzbekistan, using nine SSR markers.

MATERIALS AND METHODS Plant material

The breeding material comprised a collection of 96 samples representing four diverse species of the genus *Aegilops* L. belonging to the southwestern region of Uzbekistan. The samples' collection occurred during the route reconnaissance expeditions in 2021. The collected 96 specimens consisted of the following: 1) 23 (1-23) belonged to *Aegilops* L. species *Ae. tauschii*, 2) 11 (24-34) from *Ae. crassa*, 3) 28 (35-62) from *Ae. cylindrica*, and 4) 34 (63-96) from the species *Ae. triuncialis*. Details of collection points are available in Figure 1 and Table 1.

DNA isolation

Total genomic DNA isolation from the tissues of fresh leaves of 10–15-day-old seedlings used the CTAB method with minor modifications (Saghai-Maroof, 1984). Using an ultraviolet photometer measured the quantity and quality of the DNA. The study utilized nine pairs of SSR markers (generated by Bio-Basic Canada Inc., China). For polymorphism analysis, evaluation engaged the genus *Aegilops* L. accessions (Table 2).

PCR analysis

The reaction medium for SSR-amplification with a volume of 25 μ l included 0.2 mM of each dNTP (Syntol, Russia), 250 µM of each primer, 1.5 mM MgCl₂ (Syntol, Russia), 10 mM Tris-HC1 (pH = 9.0), 1 unit of Taq-polymerase (Syntol, Russia), and 50-100 ng of the researched DNA. Amplification continued with the following mode: initial denaturation of double-stranded DNA - 3 min at 94 °C; 35 cycles: 94 °C - 30 s, annealing - 1 min at 50 °C, 55 °C, or 60 °C (annealing temperature depending on the SSR primers used in the analysis), elongation - 2 min at 72 °C, and the final stage - elongation 10 min at the temperature of 72 °C (Abugalieva and Turuspekov, 2009; Abugalieva et al., 2010; Sultanov et al., 2022). The PCR ran on a BioRad thermal cycler (BioRad Laboratories Inc., Hercules, California, USA).



Figure 1. Map of the route-reconnaissance survey to collect a sample of seeds of species of the genus *Aegilops* (numbers of collection points correspond to those presented in Table 1).

Collection place	Accessions	Geographical coordinates			
•	Accessions	Latitude	Longitude		
Ae. tauschii, 2n=14 (D)					
Samarkand region Samarkand district. Agalyk village, h=870	1-4	39°55′04.76″	66°89′66.49′		
amarkand region Samarkand district. Akbuyra village, h=850	5	39°51′22.79″	66°88′62.89′		
amarkand region Urgut district. Kyzylbash village, h=1121	6	39°23′17.03″	67°00′19.37′		
amarkand region Urgut district. Tersak village, h=1121	7-10	39°36′92.53″	66°94′38.18′		
amarkand region Urgut district. Amankutan village, h=1320	11	39°18′24.29″	66°55′49.11′		
ashkadarya region Kitab district. Varganza village, h=855	12	39°19′72.08″	66°98′36.83′		
ashkadarya region Kitab district. Dzhauz village, h=1225	13-14	39°11′46.08″	67°16′83.92′		
ashkadarya region Kitab district. Kitab geological reserve, h=1375	15-18	39°11′23.74″	67°17′35.27′		
ashkadarya region Kitab district. Panji village, h=719	19	39°14′80.68″	66°96′07.64′		
ashkadarya region Kitab district. Khazraty Bashi village, h=874	20-23	39°23′38.40″	67°03′65.46′		
e. crassa, 2n=42 (DDM)	24.25	20055/04 76/	66000/66 40		
amarkand region Samarkand district. Agalyk village, h=870.	24-25	39°55′04.76″	66°89′66.49′		
amarkand region Samarkand district. Akbuyra village, h=850	26	39°51′22.79″	66°88′62.89		
amarkand region Urgut district. Kyzylbash village, h=1121	27	39°23′17.03″	67°00′19.37′		
amarkand region Urgut district. Tersak village, h=1121	28-29	39°36′92.53″	66°94′38.18		
amarkand region Urgut district. Amankutan village, h=1320	30	39°18′24.29″	66°55′49.11		
ashkadarya region Kitab district. Varganza village, h=855	31	39°19′72.08″	66°98′36.83		
ashkadarya region Kitab district. Kitab geological reserve, h=1375	32	39°11′23.74″	67°17′35.27		
ashkadarya region Kitab district. Panji village, h=719	33	39°14′80.68″	66°96′07.64		
ashkadarya region Kitab district. Khazraty Bashi village, h=874	34	39°23′38.40″	67°03′65.46		
e. cylindrica, 2n=28 (CD) amarkand region Samarkand district. Agalyk village, h=870.	35-37	39°55′04.76″	66°89′66.49		
amarkand region Samarkand district. Akbuyra village, h=850	38	39°51′22.79″	66°88′62.89		
5 , 5 ,	39	39°23′17.03″	67°00′19.37		
amarkand region Urgut district. Kyzylbash village, h=1121 amarkand region Urgut district. Tersak village, h=1121	40-43	39°36′92.53″	66°94′38.18		
amarkand region Urgut district. Amankutan village, h=1121	40-43	39°18′24.29″	66°55′49.11		
5 5	44	39°18′24.29 39°19′72.08″	66°98′36.83		
ashkadarya region Kitab district, Besh-Kal village, h=785		39°19'72.08"			
ashkadarya region Kitab district. Varganza village, h=855	46-49		66°98′36.83		
ashkadarya region Kitab district. Dzhauz village, h=1225	50-51	39°11′46.08″	67°16′83.92		
ashkadarya region Kitab district, Kitab geological reserve, h=1375	52-55	39°11′23.74″	67°17′35.27		
ashkadarya region Kitab district surrounding Kitab town, h=770	56	39°19′65.25″	66°90′68.54		
Cashkadarya region Kitab district. Kuktash village, h=915	57	39°17′71.26″	67°07′55.20		
ashkadarya region Kitab district. Palandara village, h=815	58	39°16′22.07″	66°98′77.85		
ashkadarya region Kitab district. Panji village, h=719	59	39°14′80.68″	66°96′07.64		
ashkadarya region Kitab district. Khazraty bashi village, h=874 e. triuncialis, 2n=28 (UC)	60-62	39°23′38.40″	67°03′65.46		
amarkand region Samarkand district. Agalyk village, h=870.	63-66	39°55′04.76″	66°89′66.49		
amarkand region Samarkand district. Akbuyra village, h=850	67-68	39°51′22.79″	66°88′62.89		
amarkand region Urgut district. Kyzylbash village, h=1121	69	39°23′17.03″	67°00′19.37		
amarkand region Urgut district. Tersak village, h=1121	70-71	39°36′92.53″	66°94′38.18		
amarkand region Urgut district. Amankutan village, h=1320	72	39°18′24.29″	66°55′49.11		
amarkand region Orgat district. Taxtakaracha village, h=1020	72	39°28′48.88″	65°82′25.25		
amarkand region orgat district the border between Kitab $h=1620$	74	39°28′48.88″	65°82′25.25		
	75	39°28′48.88″	65°82′25.25		
amarkand region Urgut district. Sharshara h=1610	76	39°28'48.88" 39°19'72.08"	66°98′36.83		
ashkadarya region Kitab district. Besh-Kal village, h=785	76 77-78				
ashkadarya region Kitab district. Varganza village, h=855		39°19′72.08″	66°98′36.83		
ashkadarya region Kitab district. Dzhauz village, h=1225	79-82	39°11′46.08″	67°16′83.92		
ashkadarya region Kitab district. Kitab geological reserve, h=1375	83-85	39°11′23.74″	67°17′35.27		
ashkadarya region Kitab district. Karabulak village, h=735	86	39°23′38.40″	67°03′65.46		
ashkadarya region Kitab district, surroundings Kitab town, h=770	87	39°19′65.25″	66°90′68.54		
ashkadarya region Kitab district. Kuktash village, h=915	88	39°17′71.26″	67°07′55.20		
ashkadarya region Kitab district. Palandara village, h=815	89	39°16′22.07″	66°98′77.85		
ashkadarya region Kitab district. Panji village, h=719	90	39°14′80.68″	66°96′07.64		
ashkadarya region Kitab district. Khazraty Bashi village, h=874	91-93	39°23′38.40″	67°03′65.46		
ashkadarya region Shakhrisabz district. Khisorak village, h=820	94	38°53′51.11″	67°15′42.82		
ashkadarya region Shakhrisabz district. Xitoy village, h=850	95	38°52′29.21″	67°18′24.21		
ashkadarya region Shakhrisabz district. Olmali village, h=790	96	38°87′35.35″	67°30'89.28		

Table 1. Genus Aegilops accessions belonging to four local species used for the analysis

Primers	Forward and reverse direction (5'-3')	Annealing temperature °C	Base pair length	Chromosome
WSP006	F: CGTATCACCTCCTAGCTAAACTAG	55	196	4B
	R: AGCCTTATCATGACCCTACCTT			
WSP044	F: GTTGAGCTTTTCAGTTCGGC	60	176	7B
	R: ACTGGCATCCACTGAGCTG			
WSP107	F: ATTAATACCTGAGGGAGGTGC	60	188	4B
	R: GGTCTCAGGAGCAAGAACAC			
WSP130	F: AGCTCTGCTTCACGAGGAAG	60	121	7A
	R: CTCCTCTTTATATCGCGTCCC			
WSP156	F: CCAACCGTGCTATTAGTCATTC	60	279	5A
	R: CAATGCAGGCCCTCCTAAC			
WSP190	F: GTGCTTGCTGAGCTATGAGTC	60	253	5D
	R: GTGCCACGTGGTACCTTTG			
WSP192	F: GGTTTTCTTTCAGATTGCG	60	232	5D
	R: CGTTGTCTAATCTTGCCTTGC			
WSP325	F: TTTCTTCTGTCGTTCTCTTCCC	60	138	6D
	R: TTTTTACGCGTGAACGACG			
WSP513	F: ATCCGTAGCACCTACTGGTCA	60	146	4B

Table 2. Characterization of nine SSR markers used to assess the genetic diversity of 96 *Aegilops* landraces.

Fractionation of amplification products proceeded by electrophoresis in 6% denaturing polyacrylamide gel (PAAG), followed by staining in ethidium bromide visualized under ultraviolet using a 30 cm Sequi-Gen GT Sequencing Cell gel apparatus (BioRad Laboratories Inc., Hercules, California, USA).

Statistical analysis

Indicators of the genetic diversity assessment, i.e., the total number of alleles (Na), expected heterozygosity (He), observed heterozygosity (Ho), and polymorphism (PIC), used the PowerMarker 3.51 program (Liu and Muse, 2005). PowerMarker software also helped formulate the allele frequencies. Cluster analysis and creating an unweighted neighbor union tree also employed the DARwin 6.0 software package (Gascuel, 1997; Perrier and Jacquemoud-Collet, 2006). The statistical program also served various methods for determining genetic distances and genetic similarity according to Nei (PopGene32, UPGMA Unweighted Pair Group Method with Arithmetic mean). Based on the binary matrices in the PAST 3.16 program, the Dice genetic similarity coefficients among the samples attained calculations (Hammer et al., 2001).

RESULTS

SSR marker polymorphism

The use of methods of monolocus analysis and SSR-marking, most commonly used, helped to characterize and compare the levels of genetic diversity of the *Aegilops* collection comprising four local species. Selecting 96 genotypes for the analysis represented 21 places of expeditionary collections located in three agricultural regions of the southwestern region of Uzbekistan. The selection of genotypes continued based on their representation of the entire initial diversity in terms of origin and botanical species.

The result of SSR-analysis of 96 Aegilops genotypes identified the 102 fragments, 65 (63.7%)wherein were polymorphic, and 37 (36.3%) were monomorphic. The number of DNA fragments amplified by one primer ranged from 7 (WSP107) to 17 (WSP513). Using the primer WSP513 obtained the maximum number of polymorphic fragments, whereas the minimum by primer WSP107. Figure 2 shows the results of a fragment analysis of four species of the genus Aegilops applying the SSR-primer WSP190. The SSR spectrum of samples of Aegilops species obtained used the primer

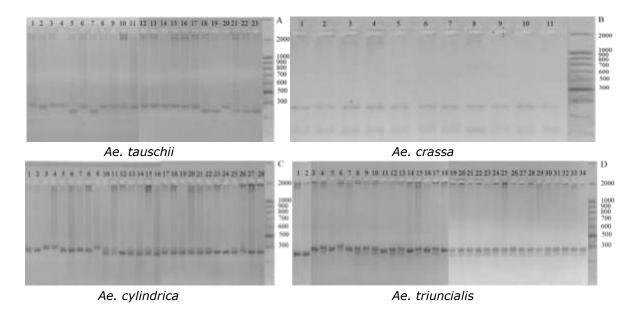


Figure 2. The SSR spectrum of the landraces of *Aegilops* species obtained using primer WSP190 (gel fragment). The rightmost lane is the molecular weight marker.

WSP190. It can also be apparent in the data that each species had different characteristics and non-repeating PCR fragments.

The SSR spectrum, using the primer WSP190 for species Ae. tauschii, Ae. crassa, Ae. cylindrica, and Ae. triuncialis, collected from various geographic locations in the southwestern region of Uzbekistan, showed inter- and intraspecific DNA polymorphism. Among the studied species, the most polymorphic were the genotypes of the species Ae. tauschii, with the SSR-primer WSP190 identifying six PCR fragments in 23 accessions. It indicates that 75% of the landraces of this species turned out to be polymorphic for this primer (Figure 2-A). In 11 accessions of the species Ae. crassa, one monomorphic fragment appeared (Figure 2-B). In addition, the use of primer WSP190 in 28 populations of the species Ae. cylindrica also identified one monomorphic portion (Figure 2-C). For 34 accessions of the species Ae. triuncialis, three PCR fragments emerged (Figure 2-D). Unlike other species, the species Ae. cylindrica and Ae. triuncialis by SSR primers revealed the lowest level of genetic variability by holding the minimum polymorphism (57.1%), respectively.

Thus, using nine SSR primers revealed a clear intraspecific difference in all the studied species of the genus Aegilops. One must also note that primers WSP006 and WSP044 in the assessed Aegilops collection have only two fragments each, and primers WSP130, WSP156, and WSP192 for individual genotypes and species owned three portions each. Identifying more than three PCR fragments surfaced in primers WSP192 and WSP513 per species of the genus Aegilops (Table 3). For nine SSR primers, 75% polymorphism was evident in the species Ae. tauschii, which might be related to its adaptive capabilities. Interestingly, the results obtained for the species Ae. crassa utterly differed from the data obtained from other species. It might be due to this species being less variable, regardless of the growth place.

The PCR fragments in different species of the genus *Aegilops* obtained using nine pairs of SSR primers appear in Table 3. As seen from the data, genotypes of the species *Ae. cylindrica* with primers WSP006, WSP107, and WSP190, and in the species *Ae. triuncialis* with primer WSP107 were 100% monomorphic. However, the species *Ae. crassa* revealed a

		Genus/species															
		Ae.	tausc	hii	Ae.	crass	а	Ae.	cylina	lrica	Ae.	triunc	cialis	_			
Locus	<i>Na, Number allelic</i>	Total	polymorph	monomorph	Total	polymorph	monomorph	Total	polymorph	monomorph	Total	polymorph	monomorph	– Ho	Н	PIC	Ι
WSP006	8	2	1	1	3	2	1	1	0	1	2	1	1	0.004	0.69	0.63	1.27
WSP044	8	3	2	1	0	0	2	3	2	1	2	2	0	0.601	0.74	0.70	1.37
WSP107	7	3	2	1	1	0	1	2	0	2	1	0	1	0.000	0.75	0.69	1.40
WSP130	13	4	3	1	5	5	0	2	1	1	2	1	1	1.000	0.68	0.64	1.28
WSP156	12	3	2	1	5	3	2	2	1	1	2	1	1	1.508	0.72	0.67	1.32
WSP190	13	5	4	1	1	0	1	2	0	2	5	1	4	1.012	0.73	0.68	1.35
WSP192	13	4	3	1	2	0	2	5	5	0	2	2	0	0.003	0.72	0.67	1.32
WSP325	11	2	1	1	5	5	0	2	1	1	2	1	1	0.010	0.73	0.68	1.35
WSP513	17	6	6	0	6	2	4	2	2	0	3	3	0	0.602	0.75	0.71	1.39
Total alleles	102													4.74	6.51	6.07	12.05
Average	11.33													0.527	0.72	0.67	1.34

Table 3. PCR amplification and genetic parameters of SSR markers in the four species of genus *Aegilops.*

low level of genetic variability, i.e., 60.7% polymorphic and 39.3% monomorphic loci, which indicates an average level of intraspecific variability according to the SSR analysis. Unlike other species, the accessions of the species *Ae. cylindrica* collected in the Samarkand region indicated having 100% polymorphic loci with primer WSP192.

In these species, the highest level of diversity may correlate with plasticity and a wide range of intraspecific variability in different habitats. The microsatellite analysis also showed that the SSR primers WSP190 and WSP192 have fragments with 800, 1000, and 1200 bp and were not polymorphic, but sections with a length of 380, 500, 750, 770, and above 1000 bp turned out to be polymorphic. The primers WSP192 and WSP513 exhibited 100% polymorphism. It is also visible from the data presented in Table 3 that the species Ae. triuncialis recorded with 57.1% of polymorphic loci and in two other species, i.e., Ae. tauschii and Ae. crassa recorded 70% and 60.7% polymorphic loci, respectively. The results indicated that the level of variability in Aegilops species with Dgenome was approximately the same.

PCR amplification and genetic diversity

Using nine SSR markers, generally amplified 102 alleles (Table 3). The number of alleles (Na) in the total sample ranged from seven (WSP107) to 12 (WSP513) and averaged 11.33 alleles per locus. The interval of expected heterozygosity (Ho) ranged from 0.000 (WSP107) to 1.508 (WSP156) and averaged 0.527. The Nei genetic diversity index (H) ranged from 0.68 (WSP130) to 0.75 (WSP107, WSP513), with an average value of 0.72. The most genetic diversity emerged for primers WSP044 and WSP513, whereas the least for WSP130 and WSP006. The PIC values of each marker locus ranged from 0.63 (WSP006) to 0.71 (WSP513) and averaged 0.68. The highest PIC value resulted from the primer WSP513 locus and the least for primer WSP006. At the same time, all the microsatellite loci had a PIC value above 0.6. The Shannon diversity index (I) in the total sample ranged from 1.27 (WSP006) to 1.40 (WSP107) and averaged 1.34.

The alleles' diversity analysis of nine microsatellite loci showed that all SSR markers produced species-specific amplicons and

	Ae. tauschii	Ae. crassa	Ae. cylindrica	
Ae. crassa	0.616			
Ae. cylindrica	0.608	0.768		
Ae. triuncialis	0.566	0.762	0.480	

Table 4. Nei's genetic distance indices between species of the genus Aegilops based on SSR analysis.

identified 35 species-specific amplicons. At the same time, the most number of alleles (15) resulted in the species *Ae. tauschii*, followed by 12 in *Ae. crassa*, eight in *Ae. cylindrica*, and five in the species *Ae. triuncialis*. However, it is noteworthy that the species *Ae. triuncialis* for SSR marker WSP044 differed by two independent alleles. Meanwhile, the primer WSP044 had the highest number of alleles and distinguished the three species, but it did not produce amplification products in the species *Ae. crassa* (Table 3).

Cluster analysis

The genetic distance (GD) index for SSR markers among all the paired combinations of samples varied from zero to 0.93, with an average value of 0.79. The study obtained the highest distance value among the genotypes 2 (*Ae. tauschii*), 38 (*Ae. cylindrica*), and 64 (*Ae. triuncialis*). However, the obtained smallest value was among various pairs of genotypes of the species *Ae. crassa*. Among the studied species, the genetic similarity was prominent between the two species *Ae. crassa* and *Ae. cylindrica* (GD = 0.77), while least between the species *Ae. cylindrica* and *Ae. triuncialis* (GD = 0.48) (Table 4).

A dendrogram constructed from the data of nine SSR markers based on the UPGMA method grouped 96 genotypes into 10 clusters (Figure 3). All four *Aegilops* species studied were significantly distinguishable and formed distinct clusters and subclusters (Figure 3). Based on cluster analysis (Figure 3-A), 23 genotypes of the species *Ae. tauschii* attained dividing into three groups (I-III). Cluster group I included genotype numbers 1-4 (Agalyk population), number 5 (Akbuyra population) of Samarkand region, and numbers 6-11 (Tersak, Kyzylbash, and Amankutan population) of District Urgut, Uzbekistan. Group II cluster

included the genotype numbers 12-23 from the District Kitab, Uzbekistan (Table 1).

Apparently, due to the specific clustering of the species Ae. tauschii by ecological geographical origins, and а conclusion may indicate the results of discriminatory analysis based on the use of SSR markers relatively accurately reflect the levels of genetic similarity of the genotypes belonging to this species and collected from different places in the southwestern region of Uzbekistan. However, one must note that the environments of the studied species showed the highest genetic diversity in the species Ae. tauschii, which characteristically indicates 75% polymorphism of this species (Table 3).

The genetic similarity analysis showed that among the genotypes of the species Ae. tauschii collected from various places in the southwestern region of Uzbekistan, no significant clustering occurred according to the location of their growth, as evidenced by the uniform polymorphism of this species. Genetic similarity analysis calculations based on SSR markers for the species Ae. crassa revealed the relative characteristics of differentiation. The genetic distance (GD) index by SSR markers among the genotypes of the species Ae. crassa ranged from 0 to 0.80, with an average value of 0.74. The highest distance value obtained emerged between genotype 27 (GD = 0.78), while the lowest genetic similarity (0.68) was among various pairs of genotypes from the District Kitab (Table 4). Based on the cluster analysis, the 11 genotypes of the species Ae. crassa gained division into different groups (Figure 3-B). The first cluster group I included genotype numbers 24-26 (population Agalyk, Akbuyra) of the District Samarkand and numbers 27-30 (populations of Kyzylbash, Amankutan) of the District Urgut. Group II clusters included genotype numbers 31-34 (populations of Varganza, Khazraty Bashi, and Panji) of the District Kitab (Table 1).

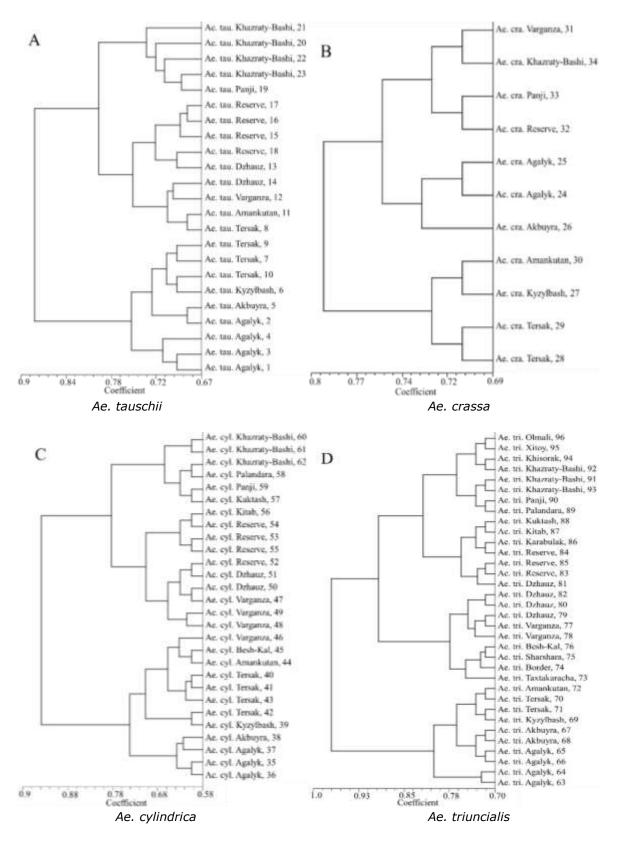


Figure 3. Dendrogram of 96 accessions of four native species of the genus *Aegilops* based on nine SSR markers.

Cluster analysis revealed a closer genetic relationship occurred among some populations from the Districts Urgut and Kitab, which were also neighboring provinces; hence, assumption for existing genetic an an said relationship among the districts' populations. It may be due to these agricultural areas' genetic changes associated with soil and climatic conditions. Similarly, the SSR analysis showed that among all the studied genotypes of the species Ae. crassa has a lower level of intraspecific diversity (60.7% of polymorphic loci), indicating an average level of intraspecific variability of this species (Table 3).

The dendrogram obtained from nine SSR markers made it possible to isolate the species Ae. cylindrica into two clusters, within which the samples have the highest genetic similarity. Based on molecular genotyping for nine SSR markers an outcome showed relatively higher genetic diversity in the species Ae. cylindrica, which was consistent with previous findings by Moradkhani et al. (2015) and Pour-Aboughadareh et al. (2017). The genetic similarity analysis based on SSR markers revealed a high level of intraspecific variability for the genotypes of the species Ae cylindrica populations. The maximum intraspecific similarity in this species was 0.90, whereas the minimum was 0.58, with an average value of 0.76 (Table 4). The farthest distance value resulted in genotype number 39 (GD = 0.89), with the smallest (0.68) among the genotype pairs collected from the District Kitab (Table 4).

The species *Ae. cylindrica*, represented in the collection by 28 samples from the Samarkand, District Urgut of the Samarkand region, and the District Kitab of the Kashkadarya region, were divided into two groups, within which the genotypes have a substantial genetic similarity. The first cluster group I included 10 genotype numbers 35-38 (populations of Agalyk and Akbuyra) and numbers 39-44 (populations of Kyzylbash, Tersak, and Amankutan) of the Districts Urgut and Samarkand. However, the second cluster, further subdividing into two subclusters, grouped 18 accessions (populations from Varganza, Khazraty Bashi, and Dzhauz) of the District Kitab.

Cluster analysis showed that populations from the same agricultural area also combined in the same group, and their further division into groups showed diversity. Such differentiation of species *Ae. cylindrica* generally corresponds to the geographical origin of the accessions. The populations most distant from each other (Districts Kitab and Samarkand) formed separate groups, and the populations closest to each other of the Districts Samarkand and Urgut came together into one group.

Based on molecular genotyping by SSR primers, results showed different degrees of intra- and interspecies polymorphism in the species *Ae. triuncialis* and *Ae. triuncialis* (Figure 3-D). The said polymorphism was evident in the collection of 34 landraces from 21 diverse locations in the southwestern region of Uzbekistan and characteristic of a significant intraspecific diversity level. Species *Ae. triuncialis* appeared as one common species in various agricultural zones of Southwestern Uzbekistan.

Based on the coefficient of genetic similarity, the species Ae. triuncialis sustained division into four groups (I-IV). The first cluster included 10 accessions (collected from Agalyk and Akbuyra) of the District Samarkand and the populations of Kyzylbash, Tersak, and Amankutan of the District Urgut. The second cluster included nine accessions (collected from Varganz and Besh-Kal). The third cluster contained seven accessions (collected from Karabulak and Kuktash). The fourth cluster comprised eight samples (collected from Khazraty Bashi, Panji, and Palandara) of the District Kitab. The maximum value of the Dice genetic similarity index among all the accessions of the species Ae. triuncialis was 1.00 (observed for sample number 13 [Agalyk population from the District Samarkand]), while the minimum was 0.89 (sample number 87 from District Kitab). According to the SSR analysis, the Dice genetic similarity index values among the accessions of the entire sample were higher than for an individual population.

DISCUSSION

Wheat's wild relatives are potential sources of valuable genetic material for its improvement. Among the wild relatives, Aegilops is the largest genus of the tribe Triticeae, which includes 23 annual species with diverse levels of ploidy and are promising sources of economically valuable traits that can benefit wheat improvement programs (Reynolds et al., 2012; Tuler et al., 2015; Yang et al., 2018; Kishii, 2019). The SSR markers have wide usage to assess the genetic diversity of genus Aegilops species and to purposefully use its pool the breeding gene in process (Noormohammadi et al., 2014; Tang et al., 2015; Henkrar et al., 2016; Liu et al., 2016; Luo et al., 2017; Pandian et al., 2018; Itam et al., 2020). These research works emphasized the role of introducing wild relative genes into the existing crop cultivars that are of great value for global food security (Brozynska et al., 2015; Redden, 2015; Buronov et al., 2023).

Genetic diversity studies of genus Aegilops species using molecular genetics tools based on the analysis of DNA polymorphism (RAPD, AFLP, SSR, and SNP) allow obtaining an individual characteristic of a separate genotype - a DNA profile. Based on long-term studies of the genetic diversity of 20 samples of five unique species of the genus Aegilops-Triticum using 10 SSR primers, findings showed a higher number of alleles per locus (Moradkhani et al., 2015). In addition, Naghavi et al. (2009) assessed the genetic diversity of 52 genotypes of diploid wheat Tr. boeoticum, using 21 microsatellite markers, and the average number of alleles per locus was 13.0 (varying from three to seven). In the existing genetic diversity study of four species of the genus Aegilops, i.e., Ae. tauschii, Ae. crassa, Ae. cylindrica, and Ae. triuncialis using nine pairs of SSR markers, the number of alleles (Na) in the total sample ranged from four (WSP107) to 12 (WSP006) and averaged 7.78 alleles per locus, which is consistent with the findings of Moradkhani et al. (2015). Saeidi et al. (2006) assessed the genetic diversity of Iranian populations of Ae. tauschii using 13 microsatellite markers, with 66 alleles amplified with a mean PIC of 0.65.

In the presented study, by assessing 96 accessions of four diverse species of the genus Aegilops, the maximum PIC value was 0.70 with a mean value of 0.68, which was 0.03 more than the value reported by Naghavi et al. (2009). Similarly, the genetic diversity of 46 Commelina communis populations using 12 SSR markers gained assessing with a mean PIC value of 0.20 (Yang et al., 2018). Pour-Aboughadareh et al. (2017) and Roder et al. (1995) also reported the variability of the PIC values that occurred dependent on the content (GT), the number of alleles per locus, and the type of motifs. In the pertinent study, nine SSR markers used comprised four microsatellite loci primers, viz., WSP006, WSP044, WSP130, and WSP513 that were the most polymorphic, which can help identify the species Ae. tauschii, Ae. triuncialis, and Ae. crassa. The SSR marker WSP006 revealed 67% of polymorphic fragments (Ae. triuncialis), 80% (Ae. tauschii), and 67% (Ae. crassa), as well as, 75% with primer WSP130 that were either species-specific or accession specific, to discriminate the two and three species.

Despite the different numbers of studied samples for each species, research further explored the genetic diversity at the species level. In this study, species Ae. triuncialis, represented by 34 accessions, had a higher level of genetic diversity. The species Ae. tauschii, represented by 23 samples, the third species in terms of volume, was characteristic of the highest level of genetic diversity. The extensive variety of the D genome in the species Ae. tauschii had confirmation with AFLP markers (Dvorak et al., 1998; Lelley et al., 2000). As for the polyploid species, Ae. crassa and Ae. cylindrica, with a D genome, also have a lower level of intraspecific diversity. The low disparity of the species Ae. cylindrica was consistent with previous findings by Goryunova et al. (2004). In contrast to the latest results, Moradkhani et al. (2015) and Pour-Aboughadareh et al. (2017) identified a relatively higher diversity of the species Ae. cylindrica compared with other species based on SSR and SCoT markers. The D-genomic groups of the plants had less intraspecific polymorphism characterization than most polyploid species with a U genome. The RAPD

analysis showed no difference between the ploidy of two species of the genus *Aegilops;* however, these species were very similar morphologically, with some researchers not even considering them as two independent species (Goryunova *et al.*, 2004).

The promising results showed that accessions of the species Ae. crassa have a closer level of genetic similarity to Ae. tauschii than with Ae. cylindrica, which seems related to the fact that Ae. cylindrica, in addition to the D genome, also has a C genome. The polyploid species Ae. triuncialis does not have a D genome but has a C genome and a U genome, wherein this species forms a single cluster with Ae. cylindrica, which also appeared in the dendrogram of this relevant study. There is also evidence in the literature that the species Ae, cvlindrica and Ae, triuncialis have a mutual C genome, confirmed by the presence of a shared cluster according to RAPD analysis (Kavrakova, 2009). According to recent results, in contrast to the species with the D genome, representatives of the species with the U genome (Ae. triuncialis) were characteristic of significant level а of intraspecific diversity. The species Ae. triuncialis proved one of the most widespread with a high level of genetic diversity. Clustering based on genetic distance was consistent with the taxonomy. The dendrogram obtained from nine pairs of SSR markers made it possible to identify all the studied species of the genus Aegilops at the level of a cluster or subcluster, which proved consistent with the findings of Pour-Aboughadareh et al. (2017).

All the accessions of the *Aegilops'* four species sustained division into three groups, i.e., a) genotypes collected from Agalyk and Akbuyra, with both of these points included in the foothill zones of the Samarkand region, b) genotypes from the Urgut mountainous district, and c) genotypes collected from various agricultural centers of the District Kitab. The District Kitab is geographically adjacent to the Samarkand region and has locations in the eastern part of the Kashkadarya region on the spurs of the Zeravshan and Gissar ranges. The main territory of these districts lies on the Kitab-Shakhrisabz lowland. The altitudinal boundary here has a contrasting presentation from the lowest (700 masl) to the highest (2000 masl) farming zones.

Two trends can be distinct in the genetic diversity of local species of the genus Aegilops. Firstly, the use of SSR markers to identify phylogenetic relationships among the species and intraspecific polymorphism of representatives of the genus Aegilops in the flora of the Districts Samarkand, Urgut, and Kitab, Uzbekistan, showed variations among the genotypes. It may be due to differences in soil and climatic conditions in the areas where the landraces were collected. However, all districts are characteristic of three а continental arid climate. The second trend was the joint grouping of the genotypes collected from Samarkand and Urgut. An analysis of the influence of longitude, latitude, and height of sampling districts on clumping within clusters showed that these parameters do not affect the genetic diversity of the studied collection. Consequently, the latitudes and heights above sea level of the Districts Samarkand, Urgut, and Kitab vastly differed. Moghaddam et al. (2000, 2014) reported the genetic diversity of Triticum-Aegilops and Triticum urartu Thum using SSR, ISSR markers, and isoenzymes, those similar geographically distant regions in terms of their conditions can lead to a joint grouping of geographically distant populations.

According to current results, the grouping of accessions of different species of the genus Aegilops from varied agricultural zones of the southwestern region of Uzbekistan according to SSR markers occurred by species, subspecies, areas, longitude, latitude, and altitude, revealing no influence on the intraspecific grouping of landraces. For more information, clustering samples from the same agricultural area continued into the same subcluster. Grouping analysis showed that within the same district, genetic parameters closely correlate to the geographical origin of the samples, and the joint grouping of specimens from different areas within the region was most likely due to the distribution of populations in similar climatic conditions. As a rule, the populations of the Samarkand region showed a distant genetic relationship with populations from the Districts Urgut and Kitab, which might refer to long-term evolution

and genetic variations associated with local climatic conditions. As a result of SSR marking, estimating the genetic diversity of local populations of four species of the genus from Southwestern Aegilops originating Uzbekistan ensued, establishing the degree of their polymorphism and genetic diversity in of the allelic composition terms of microsatellite loci.

CONCLUSIONS

Central Asia is one of the centers of origin of the genus Aegilops species. Uzbekistan also lies in this region, with the southwestern zone mainly mountainous and has a unique plant genetic treasure. Thus, a collection comprising 96 accessions of four local species of the genus Aegilops ensued from 21 expedition sites located in four agricultural regions, mostly mountainous in Southwestern Uzbekistan. As a result of SSR marking, determining the degree of genetic diversity and similarity of the genomes of these landraces for the first time also attained evaluation. The degree of polymorphism and genetic diversity of microsatellite loci collected bv allelic composition was evident. The gene pool of native species of the genus Aegilops from the southwestern region of Uzbekistan can provide valuable alleles for wheat improvement and adaptation programs.

ACKNOWLEDGMENTS

The authors are grateful to the Institute of Genetics and Plants Experimental Biology, Academy of Sciences of Uzbekistan, for the support of scientific research.

REFERENCES

Abbas A, Yu HY, Cui HL, Yu HL, Li XJ (2020). Assessment of the genetic diversity in *Aegilops tauschii* (Coss.) by using SSR markers and morpho-physiological traits. *Appl. Ecol. Environ. Res.* 7011–7020. http://doi.org/10.15666/aeer/1805_701170 20.

- Abbasov M, Akparov Z, Gross T, Babayeva S, Izzatullayeva V, Hajiye E, Rustamov K, Gross P, Tekin M, Akar T, Chao S (2018). Genetic relationship of diploid wheat (*Triticum spp.*) species assessed by SSR markers. *Genet. Resour. Crop Evol.*:1–13.
- Abugalieva SI, Turuspekov YK (2009). DNA markers in genetics and breeding of cereal crops: Methodical recommendations. Almaty. pp. 84.
- Abugalieva SI, Volkova LA, Turuspekov YK (2010). The variation of SSR profiles in bread wheat germplasm of Kazakhstan. 8th International Wheat Conference. St. Petersburg, 2010. p. 1.
- Aliyev RT, Abbasov MA, Mammadov AC (2007). Genetic identification of diploid and tetraploid wheat species with RAPD markers. *Turk J. Biol.* 31(3): 173–180.
- Alnaddaf LM, Moualla MY, Haider N (2012). The genetic relationships among *Aegilops* L. and *Triticum* L. species. *Asian J. Agric. Sci.* 4(5): 352–367. https://www.researchgate.net/ publication/280932914.
- Arzani A, Ashraf M (2017). Cultivated ancient wheats (*Triticum* spp.): A potential source of health-beneficial food products. *Comp. Rev. Food Sci. Saf.* 16: 477–488. https://doi.org/10.1111/1541-4337.12262.
- Brozynska MA, Furtado RJ, Henry (2015). Genomics of crop wild relatives: Expanding the gene pool for crop improvement. *Plant Biotechnol. J.* 14: 1070–1085.
- Buronov A, Amanov B, Muminov Kh, Tursunova N, Umirova L (2023). Polymorphism and inheritance of gliadin proteins in wheat landraces of Uzbekistan. *SABRAO J. Breed. Genet*. 55(3): 671-680. http://doi.org/10.54910/sabrao2023.55.3.6.
- Chorshanbiev NE, Nabiev SM, Azimov AA, Shavkiev JSH, Pardaev EA, Quziboev AO (2023). Inheritance of morpho-economic traits and combining ability analysis in intraspecific hybrids of Gossypium barbadense L. SABRAO. J. Breed. Genet. 55(3): 640-652. http://doi.org/10.54910/sabrao2023.55.3.4.
- Dvorak J, Luo MC, Yang ZL (1998). Genetic evidence on the origin of *Triticum aestivum* L. In: A.B. Damania, J. Valkoun, G. Willcox, and C.O. Qualset (eds.). The Origins of Agriculture and Crop Domestication. *Proceed. Harlan Symp.* ICARDA, Aleppo, pp. 235–251.
- Ehtemam MH, Rahiminejad MR, Saeidi H, Tabatabaei BES, Krattinger SG, Keller B (2010). Relationships among the A genomes of *Triticum* L. species as evidenced by SSR

markers in Iran. Int. J. Mol. Sci. 11: 4309-4325.

- Elbashir AAE, Gorafi YSA, Tahir ISA, Kim JS, Tsujimoto H (2017). Wheat multiple synthetic derivatives: A new source for heat stress tolerance adaptive traits. *Breed. Sci.* 67: 248–256. https://doi.org/10.1270/ jsbbs.16204.
- Fufa H, Baenziger BS, Beecher BS, Graybosch RA, Eskridge KM, Nelson LA (2005). Genetic improvement trends in agronomic performances and end-use quality characteristics among hard red winter wheat cultivars in Nebraska. *Euphytica* 144(1-2): 187–198.
- Gascuel O (1997). Concerning the NJ algorithm and its unweighted version, UNJ. In: Mathematical hierarchies and biology. DIMACS workshop, series in discrete mathematics and theoretical computer science. *Am. Math. Soc.* 37:149–170.
- Goryunova SV, Kochieva EZ, Chikida NN, Pukhalskyi VA (2004). Phylogenetic relationships and intraspecific variation of D-genome *Aegilops* L. as revealed by RAPD analysis. *Russ J. Genet.* 40: 515–523.
- Hajiyev ES, Akparov ZI, Aliyev RT, Saidova SV, Izzatullayeva VI, Babayeva SM, Abbasov MA (2015). Genetic polymorphism of durum wheat (*Triticum durum* Desf.) accessions of Azerbaijan. *Russ J. Genet.* 51(9): 863–870.
- Hammer Ø, Harper DAT, Paul D, Ryan (2001). Past: Paleontological statistics software package for education and data analysis. *Palaeontologia Electr*. 4(1): 9.
- Henkrar F, El-Haddoury J, Ouabbou H, Nsarellah N, Iraqi D, Bendaou N, Udupa SM (2016). Genetic diversity reduction in improved durum wheat cultivars of Morocco as revealed by microsatellite markers. *Sci. Agric.* 73(2): 134–141.
- Itam M, Abdelrahman M, Yamasaki Y, Mega R, Gorafi Y, Akashi K, Tsujimoto H (2020). *Aegilops tauschii* introgressions improve physiobiochemical traits and metabolite plasticity in bread wheat under drought stress. *Agronomy* 10: 1–17.
- Kavrakova ZB (2009). Study of DNA polymorphism in species of the genus *Aegilops* L. growing in different natural and climatic conditions of Tajikistan. Dissertation. 31–33.
- Kurbanbaev I, Abdushukirova S, Toshmatov Z, Amanov A, Azimov A, Shavkiev J (2023). Assessment of botanical and genetic collection of soybean for morphological and yield attributes and their impact on noduleassociated bacteria and soil fertility. SABRAO J. Breed. Genet. 55(3): 760-777.

http://doi.org/10.54910/sabrao2023.55.3.1 4.

- Kilian B, Mammen K, Millet E, Sharma R, Graner A, Salamini F, Hammer K, Ozkan H (2011). Aegilops in Wild Crop Relatives: Genomic and Breeding Resources. Cereals, C. Kole (ed.) Berlin: Springer. 1–76. http://dx.doi.org/10.1007 /978-3-642-14228-4_1.
- Kishii M (2019). An update of recent use of *Aegilops* species in wheat breeding. *Front. Plant Sci.* 10: 585, https://doi.org/10.3389/ fpls.2019.00585.
- Konstantinos GT, Bebeli PJ (2010). Genetic diversity of Greek *Aegilops* species using different types of nuclear genome markers. *Mol. Phylog. Evol.* 56: 951–961.
- Lelley T, Stachel M, Grausgruber H, Vollmann J (2000). Analysis of relationships between *Aegilops tauschii* and the D genome of wheat utilizing microsatellites. *Genome* 43: 661–668.
- Liu K, Muse SV (2005). PowerMarker: Integrated analysis environment for genetic marker data. *Bioinformatics* 21: 2128–2129.
- Liu XB, Feng B, Li J, Yan C, Yang ZL (2016). Genetic diversity and breeding history of Winter Mushroom (*Flammulina velutipes*) in China uncovered by genomic SSR markers. *Gene* 591: 227–235.
- Luo M, Gu YQ, Puiu D, Wang H, Twardziok SO, Deal KR, Huo N, Zhu T, Wang L, Wang Y, McGuire PE, Liu Sh, Long H, Ramasamy RK, Rodriguez JC, Van SL, Yuan L, Wang Z, Xia Z, Xiao L, Anderson OD, Ouyang Sh, Liang Y, Zimin AV, Pertea G, Qi P, Bennetzen JL, Dai X, Dawson MW, Müller H, Kugler K, Rivarola-Duarte L, Spannagl M, Mayer KFX, Lu F, Bevan MW, Leroy P, Li P, You FM, Sun Q, Liu Z, Lyons E, Wicker T, Salzberg SL, Devos KM, Dvořák J (2017). Genome sequence of the progenitor of the wheat D genome *Aegilops tauschii. Nature*. 551: 498–502.

https://doi.org/10.1038/nature24486.

Makamov A, Shavkiev J, Kholmuradova Μ. Boygobilov U, Normamatov I, Norbekov J, Khusenov N, Kushakov SH, Yuldasheva Z, Khoshimov S, Buriev Z (2023). Cotton genotypes appraisal for morphophysiological and yield contributing traits under optimal and deficit irrigated conditions. SABRAO J. Breed. Genet. 55(1): 74-89.

http://doi.org/10.54910/sabrao2023.55.1.7.

Martynov SP, Dobrotvorskaya TV, Mitrofanova OP (2015). Genealogical analysis of the use of aegilops (*Aegilops* L.) genetic material in wheat (*Triticum aestivum* L.). *Russian J. Genet.* 51(9): 855–862.

- Masoumi SM, Kahrizi D, Rostami-Ahmadvandi H, Soorni J, Kiani S, Mostafaie A, Yari K (2012). Genetic diversity study of some medicinal plant accessions belongs to Apiaceae family based on seed storage proteins patterns. *Mol. Biol. Rep.* 39: 10361–10365.
- Moghaddam M, Ehdaie B, Waines G (2000). Genetic diversity in populations of wild diploid wheat (*Triticum urartu* Thum. ex Gandil) revealed by isozymes markers. *Genet. Resour. Crop Evol.* 47: 323–334.
- Moghaddam M, Pirbalouti AG, Mehdizadeh L, Pirmoradi MR (2014). Changes in composition and essential oil yield of *Ocimum ciliatum* at different phenological stages. *Eur. Food Res. Technol.* 240(1): 199–204.
- Moradkhani H, Mehrabi AA, Etminan A, Pour-Aboughadareh A (2015). Molecular diversity and phylogeny of *Triticum-Aegilops* species possessing D genome revealed by SSR and ISSR markers. *Plant Breed. Seed Sci.* 71: 82–95.
- Naghavi MR, Maleki M, Alizadeh H, Pirseiedi M, Mardi M (2009). An assessment of genetic diversity in wild diploid wheat *Triticum boeoticum* from West of Iran using RAPD, AFLP, and SSR markers. *J. Agric. Sci. Technol.* 11: 585–598.
- Noormohammadi Z, Trujillo I, Belaj A, Ataei S, Hosseini-Mazinan M (2014). Genetic structure of Iranian olive cultivars and their relationship with Mediterranean's cultivars revealed by SSR markers. *Sci. Hortic.* 178: 175–183.
- Ogbonnaya FC, Abdalla O, Mujeeb-Kazi A, Kazi AG, Xu SS, Gosman N, Lagudah ES, Bonnett D, Sorrells ME, Tsujimoto H (2013). Synthetic hexaploids: Harnessing species of the primary gene pool for wheat improvement. *Plant Breed. Rev.* 37: 35–122. https://doi.org/10.1002/9781118497869.ch2.
- Pandian S, Satish L, Rameshkumar R, Muthuramalingam P, Rency AS, Rathinapriya P, Ramesh M (2018). Analysis of population structure and genetic diversity in an exotic germplasm collection of *Eleusine coracana* (L) Gaertn. using genic-SSR markers. *Gene* 653: 80–90.
- Perrier X, Jacquemoud-Collet JP (2006). DARwin software. http://darwin.cirad.fr/darwin.
- Pour-Aboughadareh A, Ahmadi J, Mehrabi AA, Etminan A, Moghaddam M, Siddique KHM (2017). Physiological responses to drought stress in wild relatives of wheat:

Implications for wheat improvement. *Acta Physiol. Plant.* 39: 106. http://dx.doi.org/ 10.1007/s11738-018-2651-6.

- Redden R (2015). Wild relatives for the crop improvement challenges of climate change: The adaptation ranges of crops. In R. Redden, S.S. Yadav, N. Maxted, M.E. Dulloo, L. Guarino, and P. Smith (eds.) Crop Wild Relatives and Climate Change (pp. 61–76). New Jersey, USA: John Wiley & Sons. http://dx.doi.org/10.1002/ 9781118854396.ch4.
- Reynolds M, Foulkes J, Furbank R, Griffiths S, King J, Murchie E, Parry M, Slafer G (2012). Achieving yield gains in wheat. *Plant Cell Environ.* 35: 1799–1823. https://doi.org/ 10.1111/j.1365-3040.2012.02588.x.
- Roder MS, Plaschke J, König SU, Borner A, Sorrells ME, Tanksley SD, Ganal MW (1995). Abundance, variability and chromosomal location of microsatellites in wheat. *Mol. Gen. Genet.* 246: 327–333.
- Saeidi H, Rahiminejad MR, Vallian S, Heslop-Harison JS (2006). Biodiversity of diploid D-genome *Aegilops tauschii* Coss. in Iran measured using microsatellites. *Genet. Resour. Crop Evol.* 53: 1477–1484.
- Saghai-Maroof MA, Soliman KM, Jorgensen RA, Allard RW (1984). Ribosomal DNA spacer-length polymorphisms in barley: Mendelian inheritance, chromosomal location, and population dynamics. *Proceed. Nat. Acad. Sci.* 81(24): 8014–8018. https://doi.org/10.1073/pnas.81.24.8014.
- Shavkiev J, Nabiev S, Azimov A, Chorshanbiev N, Nurmetov KH (2022). Pima cotton (*Gossypium barbadense* L.) lines assessment for drought tolerance in Uzbekistan. *SABRAO J. Breed. Genet.* 54(3): 524–536.

http://doi.org/10.54910/sabrao2022.54.3.6.

- Shavkiev J, Azimov A, Khamdullaev S, Karimov H, Abdurasulov F, Nurmetov K (2023). Morphophysiological and yield contributing traits of cotton varieties with different tolerance to water deficit, Journal of Wildlife and Biodiversity, 7(4), 214-228.
- Sobirov F, Djabbarov I (2021). Distribution of species of the genus *Aegilops* L. in the South West of Uzbekistan. *Bull. Sci. Pract.* 7(10): 72–83.
- Sultanov A, Lee E, Park H, and Cho Y (2022). Antiinflammatory Effect of Wild Indigo (Baptisia tinctoria) Root on Raw 264.7 Cells with Stimulated Lipopolysaccharide. Horticultural Science and Technology. 40 (1): 109-119
- Tang XT, Tao HH, Du YZ (2015). Microsatellite-based analysis of the genetic structure and

diversity of *Aleurocanthus spiniferus* (Hemiptera: Aleyrodidae) from tea plants in China. *Gene* 560: 107–113.

- Tilman D, Balzer C, Hill J, Befort BL (2011). Global food demand and the sustainable intensification of agriculture. *Proc. Natl. Acad. Sci. USA* 108: 20260–20264. https://doi.org/10.1073/pnas.1116437108.
- Tuler AC, Carrijo TT, No'ia LR, Ferreira A, Peixoto AL, da Silva Ferreira MF (2015). SSR markers: A tool for species identification in Psidium (Myrtaceae). *Mol. Biol. Rep.* 42(11): 1501–1513.
- Van-Slageren MW (1994). Wild Wheats: A Monograph of *Aegilops*, L. and

Amblyopyrum (Jaub. et Spach) Eig (Poaceae). Wageningen Agricultural University, Wageningen and ICARDA, Aleppo; Wageningen. https://www.researchgate.net/publication/3 01694954.

- Vieira MLC, Santini L, Diniz AL, Munhoz CF (2016). Microsatellite markers: What they mean and why they are so useful. *Genet. Mol. Biol.* 39: 312–328.
- Yang J, Yu HY, Li XJ, Dong J (2018). Genetic diversity and population structure of *Commelina communis* in China based on simple sequence repeat markers. *J. Integr. Agric.* 17: 2292–2301.